

# Photosynthesis of seedlings of *Otoba novogranatensis* (Myristicaceae) and *Ruagea glabra* (Meliaceae) in abandoned pasture, secondary forest and plantation habitats in Costa Rica

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**Abstract:** Fotosíntesis de plántulas de *Otoba novogranatensis* (Myristicaceae) y *Ruagea glabra* (Meliaceae) en pastizales abandonados, bosques secundarios y hábitats de plantaciones en Costa Rica. Enrichment planting in naturally recovering secondary forests or in tree plantations is increasingly being used as strategy to restore later-successional, large-seeded tropical forest trees. We seeded two tree species (*Otoba novogranatensis* and *Ruagea glabra*) in three agricultural sites in Southern Costa Rica: abandoned pastures, eight to ten year old secondary forests and three year old tree plantations (containing two N-fixing of four total tree species). We measured micrometeorological conditions, soil water content, plant water potential, leaf area, foliar C and N, and photosynthesis to better understand mechanistic responses of seedlings to conditions in the different successional habitats. Micrometeorological conditions, soil water content, and plant water potential were generally similar across habitats. Certain aspects of leaves (such as Specific Leaf Area and foliar N content), and photosynthesis (e.g. quantum yield and electron transport rate) were highest in the plantations, intermediate in the secondary forests, and lowest in abandoned pastures. Enhanced rates of photosynthetic biochemistry (such as  $V_{cmax}$  and  $J_{max}$ ) and Photosystem II efficiency (e.g. thermal energy dissipation) occurred in leaves from the plantations compared to the abandoned pastures, which may be related to higher leaf %N content. Results suggest that foliar N may be of greater importance than soil water content and micrometeorological factors in driving differences in photosynthetic processes across planting habitats. Planting seeds of these two species in plantations containing three year old trees (including two N-fixing species) enhances certain aspects of their photosynthesis and growth, compared to seedlings in abandoned pastures with non-native grasses, and thus can help increase forest recovery on abandoned agricultural lands. Rev. Biol. Trop. 61 (3): 1493-1507. Epub 2013 September 01.

**Key words:** chlorophyll *a* fluorescence, forest restoration, gas exchange, leaf N content, Photosystem II, tropical secondary forest.

Efforts to restore tropical forests have increased substantially over the past decade. In some cases tropical forests regenerate quickly when agriculture is ceased, whereas in heavily-used sites located far from seed sources, recovery is often sufficiently slow that intervention is necessary to accelerate forest recovery (Lamb *et al.* 2005, Chazdon 2008). The most common strategy for restoring tropical forest is to plant a mix of relatively fast-growing tree species that establish a canopy, ameliorate stressful microclimate conditions, improve nutrient availability, shade out light

demanding grasses, and attract seed dispersers (Holl 2002, Lamb *et al.* 2005, Chazdon 2008). The assumption is that these species will facilitate the subsequent establishment of a wide suite of forest plants. An increasing number of studies, however, have shown that colonization of larger-seeded and later-successional tree species can be extremely slow, even in cases where there is remnant forest nearby (Martínez-Garza & Howe 2003, Zimmerman *et al.* 2007, Cole *et al.* 2010).

One strategy used to increase forest diversity is to “enrich” these sites by seeding or



planting later successional species once an initial tree canopy has been established (Lamb *et al.* 2005, Piotto 2007). Several studies have shown that medium- to large-seeded mature forest trees can be seeded successfully into either secondary forests or tree plantations at a relatively low cost (Engel & Parrotta 2001, Camargo *et al.* 2002, Bonilla-Moheno & Holl 2010, Cole *et al.* 2011). However, there has been relatively little consideration of how the composition of the initially planted species affects the growth of species planted in later years.

Our past research shows that seedling growth, relative allocation to above-ground biomass, and foliar N concentrations were higher in seedlings growing in the understory of mixed species plantations with two N-fixing species, compared to secondary forests or abandoned pastures in Southern Costa Rica (Cole *et al.* 2011). For example, seedling survival was 20% higher, seedling height 33% higher and seedling biomass 2-fold greater in plantations compared to secondary forests and abandoned pastures. Foliar N content was almost twice as high in the plantations as in secondary forest and abandoned pasture habitats. Moreover, litterfall N concentration was higher in plantations than secondary forests, whereas concentrations of several other nutrients (e.g. Ca, Mg, K, Zn and Mn) were higher in secondary forest litter (Celentano *et al.* 2011). Foliar P concentration was somewhat higher in the plantations and did not differ between abandoned pastures and secondary forests (Cole *et al.* 2011); surprisingly, litterfall P and C:P did not differ across the three habitats (Celentano *et al.* 2011). These results, as well as other studies (Cusack & Montagnini 2004, Siddique *et al.* 2008), suggest that the initial species can strongly influence nutrient dynamics and growth, and in turn, the successional trajectory in a site.

Most past studies of tree planting to restore tropical forest have primarily measured seedling survival and growth, despite the fact that well-replicated, manipulative restoration treatments provide an excellent opportunity

to better understand the physiological mechanisms explaining successional patterns of forest seedling dynamics and further our knowledge of tropical seedling physiology. Moreover, knowledge of the particular abiotic factors (e.g. water, light, or nutrient availability) that most limit growth can help inform tropical restoration plans (Loik & Holl 1999), and how to best allocate the limited resources available to restore the extensive areas of degraded tropical lands (Holl & Aide 2011). The physical environment may limit survival and growth of seedlings used in restoration efforts by reducing intrinsic photosynthetic capacity through stomatal limitations to carbon uptake, by reducing the efficiency of light capture and electron transport within Photosystem II (PSII), or by reducing the rate of biochemical processes associated with carbon fixation, utilization of triose phosphates, phloem loading, and sink strength (Pammenter *et al.* 1993, Raines 2003, Amiard *et al.* 2005, Minchin & Lacombe 2005, Demmig-Adams & Adams 2006, Anderson *et al.* 2008).

The goal of this study was to better understand the physiological mechanisms underlying differences in growth for seedlings direct-seeded in abandoned pastures, secondary forests, and plantations that contain two N-fixing trees to restore premontane tropical forest in Southern Costa Rica. In this study, we measured a variety of structural (e.g. leaf area, %C, %N,  $\delta^{13}\text{C}$ ,  $\delta^{15}\text{N}$ ) and physiological (e.g. water relations and photosynthesis) characteristics for two common tree species within the three habitats at three sites. We tested whether PSII efficiency, photosynthetic capacity, and biochemistry differed within the tree plantations, abandoned pasture, and secondary forest habitats.

## MATERIALS AND METHODS

**Study area:** The study was carried out in three habitats: an abandoned pasture, a secondary forest, and a plantation (see descriptions below), at three sites locally named Bambu (BB, 8°44'36" N - 82°58'04" W), Finca Loma Linda (LL, 8°44'21"N - 82°55'44" W) and

San Gabriel (SG, 8°45'28" N - 82°57'28" W), separated by >1km, and located near the town of Agua Buena, in Southern Costa Rica. Study sites ranged from 1 110 to 1 290m above sea level, with slopes of 15-35° and are located in tropical premontane rain forest (Holdridge *et al.* 1971). All pasture and plantation plots were used for a mixture of coffee and cattle pasture, the typical land-use in this region, for at least 30 years prior to cessation of active management in 2004 (Holl *et al.* 2011 Table 1 for a detailed land use history of these sites). The soils are a mix of ultisols and andisols; they are moderately acidic with high percent organic matter and relatively low P. The only soil nutrient that varied among habitat types was Ca, which was higher in plantations and secondary forests than in pastures (Cole *et al.* 2011).

**Species description:** We direct-seeded five primary forest tree species that have large seeds, as part of a study on enrichment planting (Cole *et al.* 2011). We selected two species, *Otoba novogranatensis* Moldenke (Magnoliales, Myristicaceae) and *Ruagea glabra* Triana & Planch (Sapindales, Meliaceae), that had sufficient germination and leaf area to conduct physiological measurements. Both species are locally-common, canopy trees with large, animal-dispersed, recalcitrant seeds (mean fresh seed weight - *O. novogranatensis*: 4.6±0.8g; *R. glabra*: 2.7±0.5g).

**Experimental design:** We direct-seeded into each of three habitat types: experimentally established tree plantations (three years since

tree planting); abandoned pastures (one year since grazing or clearing, hereafter referred to as 'pasture'), and secondary forests (eight to ten years since agriculture or grazing). Plantation and pasture plots (50×50m) were established in 2004 as part of a large-scale restoration research project to test the effects of different tree planting strategies on forest recovery (described in detail in Holl *et al.* 2011). We selected secondary forests that had eight to ten years of regrowth and similar previous land use to the other treatments. The study was set up as a randomized block design with site (n=3) as the blocking factor and each of the three habitats at a site located within 10-200m of each other.

The tree plantations included a mixture of four tree species: two native species with commercially valuable timber, *Terminalia amazonia* (J.F. Gmel.) Exell (Combretaceae) and *Vochysia guatemalensis* Donn. Sm. (Vochysiaceae), and two fast-growing, naturalized species, *Erythrina poeppigiana* (Walp.) O.F. Cook and *Inga edulis* Mart. (both are N-fixers in the Fabaceae), that are commonly used as shade trees in agriculture in the region. Trees were planted at a density of 1 252/ha (separated by 2.8m). At the time of direct seeding, plantations had developed closed canopies that were four to five meters tall; the canopy cover and litter-fall of the sites was almost entirely comprised of the planted species (Celentano *et al.* 2011), given that the few seedlings that had naturally recruited were quite small (<2m). Pasture plots had almost no tree canopy cover and were dominated by ruderal vegetation (1-1.5m tall)

TABLE 1  
Plant community characteristics in Pasture (Pa), Secondary Forest (Sf) and Plantation (Pl) habitats at three former agricultural sites in Southern Costa Rica

Habitat	Overstory cover (%)	Bare ground (%)	Grass cover (%)	Herbaceous cover (%)
Pa	1 (1) <sup>a</sup>	6 (10) <sup>a</sup>	56 (22) <sup>a</sup>	41 (22) <sup>a</sup>
Sf	85 (4) <sup>b</sup>	18 (24) <sup>b</sup>	33 (27) <sup>b</sup>	43 (22) <sup>a</sup>
Pl	88 (9) <sup>c</sup>	51 (31) <sup>c</sup>	9 (18) <sup>c</sup>	35 (20) <sup>a</sup>

Data are means±SD for n=three sites; means with the same superscript letter within a column are not significantly different (p<0.05 using a Tukey's HSD multiple comparison test. ANOVA values for habitat differences – Overstory cover: F=2104.1, p<0.0001; Bare ground: F=21.7, p<0.0001; Grass cover: F=23.1, p<0.0001; Herb cover: F=1.9, p=0.1224.

comprised of introduced forage grasses (primarily *Axonopus scoparius* (Flüggé) Kuhl., *Pennisetum purpureum* Schumach. and *Urochloa brizantha* (Hochst. Ex. A. Rich.) R.D. Webster); a variety of ruderal herbs (primarily in the Asteraceae) and the fern *Pteridium arachnoideum* (Kaulf.).

We marked a 50×50m plot in the center of each secondary forest (total forest size ~1-3ha). Canopy height ranged from 5-15m. The mean stem density (stems >2cm dbh/ha) for the three sites for this study were 3 698 (Bambu), 2 240 (Finca Loma Linda), and 2 708 at San Gabriel. Dominant tree species included *Cecropia obtusifolia* Bertol., *Conostegia xalapensis* (Bonpl.) D. Don ex DC., *Croton draco* Schtdl., *Heliocharpus appendiculatus* Turcz., and *Psidium guajava* L (Cole, unpublished data). We did not find any large N-fixing trees in secondary forest plots.

Within each 50×50m plot in each habitat type (abandoned pasture, plantation, and secondary forest), we established three 5×8m subplots separated by over 10m (so that seedling manipulations would not confound measurements of natural recruitment in other areas of the plot); data from the three subplots were combined. Within each subplot seeds were planted in single-species rows at a density of eight seeds per square m and a depth of ~3cm. Grasses and herbaceous vegetation in the pastures were cut immediately prior to seeding to facilitate experimental set up, but quickly grew back.

Freshly fallen seeds were collected during the peak of the fruiting season in March (*R. glabra*) and April (*O. novogranatensis*) 2007 from a minimum of four trees located in forest fragments within a ten km radius of planting sites. Seeds were mixed and stored for less than three days before being soaked for 12-24h to imbibe water. Seeds that showed signs of damage or floated in water were discarded (Schatral & Fox 1994). At the time that the current study was conducted (February-March 2008), seedlings were nine to eleven months old and ~15-25cm in height.

Seedling measurements (described below) were generally taken on five individuals in each habitat × site combination, although there were no individuals of *R. glabra* surviving in the pasture treatment at one site, so n=10-15 individuals of each species in each treatment for most measurements. Because many of the surviving seedlings in the pasture habitat were apparently in negative carbon balance at the time of the *A-Q* curves (described below), we have data for fewer individuals (n=4 or 7) in the pastures.

**Existing vegetation and meteorological conditions:** We measured existing overstory and understory vegetation cover at four points per subplot (12 points total) in March 2008. We estimated percent grass cover, forb cover, and bare ground in 1×1m quadrats using a ranking system: 0, 1-5, 5-10, 10-25, 25-50, 50-75, 75-95, and 95-100%. We used a spherical densiometer to measure canopy cover in four directions at one meter height, and averaged the values.

Rainfall and temperature data during the study period were obtained from the Organization for Tropical Studies Las Cruces Biological Station (~8km from the study sites). Micrometeorological conditions within the three habitats were monitored during the period of plant physiological measurements using LI-COR Model 1000 data loggers. A LI-COR 1000-102 soil thermistor was inserted at an angle to the surface to a depth of 1cm. A Vaisala INTERCAMP HMP integrated air temperature and relative humidity sensor was suspended within a 4cm diameter piece of pvc capped at one end, which served as a radiation shield. The radiation shield was mounted to rebar with the open end facing the soil surface so that the temperature/RH sensor was at seedling leaf height. A LI-COR LI-190 quantum sensor was mounted on top of the radiation shield to measure photosynthetic Photon Flux Density (PPFD; 400-700nm) at the height of the top of seedlings. Conditions were measured using three logger-sensor assemblies simultaneously at one randomly-selected location within each

habitat type per site for three consecutive days. Each three day period was uniform in terms of morning temperature, the timing of the onset of cloudiness, and afternoon rainfall. Although February is part of the dry season at this latitude, 2008 had a wetter-than-usual dry season.

**Soil and plant water relations:** Soil volumetric water content was measured on the days of gas exchange measurements using a HydroSense portable TDR with 20cm long probes (Campbell Scientific, Logan, UT, USA). Probes were inserted into the soil in a grid of nine locations in each plot.

Leaf water potential ( $\Psi$ ) was measured using a Scholander-type pressure chamber (PMS Instruments, Corvallis, OR, USA) during the mid-morning gas exchange measurement period on five leaves per species  $\times$  habitat  $\times$  site. Measurements were made *in situ* for leaf blades combined with ca. 5cm long petiole segments that were removed with a scissors, and then immediately inserted into the pressure chamber.

**Photosynthetic gas exchange and chlorophyll *a* fluorescence from Photosystem II:** Photosynthetic CO<sub>2</sub> assimilation (*A*) was measured from approximately 0700 to 1200 hours local time (before onset of cloudiness and daily rainfall) using a portable open-mode infra-red gas analyzer (Model 6400, LI-COR, Inc., Lincoln, NE, USA) with an integrated Leaf Chamber Fluorometer (LCF). Air temperature and vapor pressure deficit (VPD) within the leaf chamber were maintained at ambient levels. Leaf temperatures were recorded with a chromel-constantan thermocouple appressed to the abaxial surface of the leaf within the leaf chamber. The CO<sub>2</sub> concentration within the leaf measurement chamber was maintained by scrubbing the incoming airstream with soda lime, and the subsequent addition of 385 $\mu$ mol/mol of CO<sub>2</sub> via injection from an external cartridge. PFD within the chamber was maintained using the LCF Light Emitting Diodes (LEDs). For all measurements, the 2cm<sup>2</sup> leaf chamber of the LCF was used. Distal, fully

mature leaves of five randomly-selected plants of each species in each plot were inserted into the leaf chamber at their natural branch orientation by mounting the leaf chamber on a tripod. Photosynthetic measurements were recorded when all stability criteria were met when the coefficient of variation for *A* and stomatal conductance to water vapor (g<sub>s</sub>) combined was <0.5%.

In order to characterize potential stress within PSII and carbon gain in response to variation in PFD, we simultaneously measured photosynthetic gas exchange and chlorophyll *a* fluorescence light response curves across planting habitats. Upon enclosure within the chamber, each leaf was allowed to dark-adapt for 20min, followed by a measurement of the maximal quantum yield of chlorophyll *a* fluorescence from PSII in the dark-adapted state ( $F_V/F_M$ ; see next paragraph for fluorescence measurement settings). A software script that combined gas exchange and chlorophyll *a* fluorescence measurements was utilized to generate the *A-Q* (CO<sub>2</sub> assimilation vs. PFD) response and the responses of the quantum yield of PSII ( $\Phi$ PSII vs. PFD), the proportion of open PSII reaction centers (qP vs. PFD; nomenclature of Vankooten & Snel 1990), electron transport rate through PSII (ETR vs. PFD), and the engagement of energy dissipation processes within PSII (D vs. PFD). Following determination of  $F_V/F_M$ , the rate of CO<sub>2</sub> assimilation was recorded and the software script altered the fluorescence measurement and flash conditions (described below). PFD was then increased stepwise to 1 200 $\mu$ mol/m<sup>2</sup>.s, with measurements at 1 200, 700, 500, 300, 100, 60 and 0 $\mu$ mol/m<sup>2</sup>.s. PFD was measured with a gallium-arsenide-phosphide sensor within the leaf chamber. Preliminary experiments showed that stomata responded quickly (<5min) to increases in PFD. At a particular PFD level, measurements were recorded at 4s intervals, and PFD levels were changed to the next quantity when CO<sub>2</sub> uptake stabilized (three stability criteria met for CO<sub>2</sub> and H<sub>2</sub>O concentrations and flow rate, with a total CV of 0.1%).

For the determination of  $F_v/F_M$ , the measurement software was set for the LCF to operate at an intensity of one, a modulation of 0.25kHz, a filter setting of one, and a gain of ten; the saturating flash duration was 0.8s at an intensity of seven, with a modulation of 20kHz with a filter of 50kHz. For the light-adapted determination of  $\Phi_{PSII}$ , the measurement intensity was increased to five and the modulation increased to 20kHz, and the flash duration was 0.8s. For fluorescence parameters that required measurement of  $F_o'$ , PSII was oxidized by preferentially exciting Photosystem I (PSI); the actinic light source was turned off for six seconds, and the far red source intensity was set to eight. The far red source was turned on one second before the actinic turned off, and then the far red turned off one second after the actinic was turned off. The modulation was 0.25kHz, and the filter was one Hz.

Based on the  $A-Q$  curve data, dark respiration ( $R_d$ ), the apparent quantum yield ( $\Phi$ ), light compensation point (LCP), PFD at incipient saturation of  $CO_2$  assimilation ( $PFD^{sat}$ ), and maximal photosynthetic rate ( $A_{max}$ ) were estimated from each light response curve using a non-rectangular hyperbola model (Sims & Pearcy 1991). These measurements may overestimate respiration due to the gasket effect on  $CO_2$  diffusion while measuring low rates of gas exchange (Pons & Welschen 2002). We were not able to utilize bottled oxygen to minimize the Kok Effect on dark respiration.

At one site (LL), we compared photosynthetic capacity at high PFD levels by measuring rates of  $A$  in relation to varying internal leaf  $CO_2$  concentration ( $c_c$ ), or instantaneous  $A-Ci$  curves. During all measurements, PFD was held at the median daily maximum value of 1 500mmol/m<sup>2</sup>.s using the red-blue LEDs, ambient temperatures were ~18-25°C, and VPD was generally <2.50kPa. Photosynthesis was measured and  $c_c$  was calculated at each of the following leaf chamber  $pCO_2$  target values: 100, 200, 300, 400, 600, 800, 1 000 and 1 200µmol/mol. Photosynthetic capacity under saturating light and optimal ambient conditions was calculated using a non-linear

regression between  $A$  and leaf chamber  $pCO_2$ . The maximum rates of carboxylation ( $V_{cmax}$ ) and electron transport ( $J_{max}$ ), and Triose Phosphate Utilization for carbohydrate transport across the chloroplast membrane (TPU) were calculated from the  $A-Ci$  curves, and were corrected to a common temperature of 25°C (Long & Bernacchi 2003).

**Leaf structure and composition:** Leaf area (blade only) was measured using a LICOR Model LI-3100 leaf area meter. The leaves that were used for water potential measurements were returned to the laboratory, dried in a forced draft oven at 40°C for three days, and weighed. We compared the intrinsic water use efficiency ( $\delta^{13}C$ ) and potential source of N from biological fixation ( $\delta^{15}N$ ) for these leaves. Following drying, leaves were ground in a ball mill and packed in tin capsules. Percent C, %N,  $\delta^{13}C$ , and  $\delta^{15}N$  were determined on a continuous-flow elemental analyzer (Carlo Erba Model 1108, CE Elantech, Inc., Lakewood, NJ, USA) coupled to an isotope ratio mass spectrometer (ThermoFinnigan Delta Plus XP, Thermo Fisher Scientific, Waltham, MA) at the UC Santa Cruz Stable Isotope Laboratory.

We analyzed the data using a randomized-block, two-way analysis of variance (ANOVA) to compare the effects of habitat and species and their interactions on most plant response variables (e.g. fluorescence, gas exchange, isotopes, water potential) with site as the blocking factor.  $A-Ci$  curves were only conducted at a single site so they were analyzed using a two-way analysis of variance without a blocking factor.

The model residuals for most variables met assumptions of normality, but in a few cases variables were arcsine square root transformed (percentages) or log + 1 transformed to normalize data. We used Tukey's HSD test ( $p=0.05$ ) for post hoc analyses for pair-wise habitat comparisons. We report means  $\pm$  1 SE throughout. All analyses were conducted using SAS version 9.2.

## RESULTS

Overstorey tree cover was >80% in plantations and secondary forests, and non-existent in pastures (Table 1). Grass cover was highest in pastures, intermediate in secondary forests, and lowest in plantations with the reverse pattern for bare ground; herbaceous cover was similar across habitats (Table 1). Both maximum and mean PFD at seedling height (~0.20m) were highly variable across sites and habitats (Table 2), reflecting variation in understory and overstorey structure across sites. Mean air and soil temperatures and RH were relatively similar and did not show a consistent trend across habitat types (Table 2). Total precipitation for the 12 month period from seed planting to seedling physiology measurements was 4 684mm, which is 142% of the 3 294±712mm annual average for the prior 34 years. For 2008, precipitation for January through March (the dry season in this region) was 190mm (inter-annual range 167-263mm).

In general, structural, biochemical and physiological characteristics of leaves differed significantly between plantations and pastures with varying or intermediate responses in secondary forests (Table 3, Appendix). Both leaf area and specific leaf area (SLA) were highest in the plantations, intermediate in the secondary

forests, and lowest in the pastures (Table 3, Appendix); there was a significant species × habitat interaction term for leaf area, as this trend was more pronounced for *R. glabra*. Percent C and N in leaves were both significantly higher in plantation habitats planted with N-fixing overstorey trees than secondary forests and pastures, with the opposite pattern for C:N.  $\delta^{15}\text{N}$  was higher in *R. glabra*, but showed only a marginally significant habitat effect ( $F=2.8$ ,  $p=0.0651$ ) with a trend towards lower values in the plantation than the pasture sites.

Seedlings in plantations had the highest responses of photosynthetic  $\text{CO}_2$  assimilation to varying PFD for both species (Fig. 1, Table 3). Based on the light response curves, dark respiration ( $R_d$ ) was lowest (i.e. least negative) for *R. glabra* in the plantation habitats, but was similar across habitats for *O. novogranatensis* (significant habitat × species interaction). The light compensation point (LCP) was lowest in the plantation and highest in the pasture for both species, but the secondary forest average for *R. glabra* was similar to that in the plantations, whereas for *O. novogranatensis* it was similar to the pastures. Apparent quantum yield ( $\Phi$ ) was highest in plantations compared to secondary forests and pastures for *R. glabra* (the model analysis did not converge on a solution for  $\Phi$  for *O. novogranatensis*). The

TABLE 2  
Micrometeorological conditions in Pasture (Pa), Secondary Forest (Sf) and Plantation (Pl) habitats at three former agricultural sites in Southern Costa Rica<sup>1</sup>

Site	Habitat	PFD <sub>mean</sub> <sup>2</sup>	PFD <sub>max</sub>	T <sub>air</sub> <sub>mean</sub>	T <sub>air</sub> <sub>max</sub>	T <sub>soil</sub> <sub>mean</sub>	T <sub>soil</sub> <sub>max</sub>	RH <sub>mean</sub>	RH <sub>min</sub>
BB	Pa	260	1133	19	30	19	21	73	37
	Sf	67	611	20	28	19	21	80	47
	Pl	146	1574	19	35	19	23	73	26
LL	Pa	701	1701	21	36	22	27	76	37
	Sf	272	1245	19	30	20	23	80	40
	Pl	234	1038	19	33	ND <sup>3</sup>	ND	77	44
SG	Pa	58	291	19	28	19	21	77	54
	Sf	71	454	21	27	20	26	78	63
	Pl	56	147	20	25	19	21	80	53

1. Data are means or extremes (maxima or minima) measured over three consecutive days at a single point in each habitat.
2. PFD=photosynthetic Photon Flux Density (400 – 700 nm;  $\mu\text{mol}/\text{m}^2\cdot\text{s}$ ); T (°C); RH, relative humidity (%).
3. “ND” = not determined due to temporary equipment failure.

TABLE 3  
Structure, composition, and photosynthetic physiology for seedlings planted in Pasture (Pa), Secondary Forest (Sf), and Plantation (Pl) habitats<sup>1</sup>. Data are means (SD)

	<i>Otoba novogranatensis</i>			<i>Ruagaea glabra</i>			H <sup>2</sup>	S <sup>3</sup>	H × S <sup>4</sup>
	Pa	Sf	Pl	Pa	Sf	Pl			
Leaf area	36 (23)	73 (43)	178 (161)	40 (33)	138 (91)	703 (480)	Pl > Sf = Pa	***	***
SLA	181 (47)	173 (30)	217 (36)	228 (54)	269 (48)	304 (68)	Pl > Sf > Pa	***	NS
%C	44.27 (0.65)	43.96 (0.97)	45.06 (0.80)	44.86 (1.68)	45.54 (1.29)	46.64 (0.99)	Pl > Sf = Pa	***	NS
%N	0.98 (0.20)	1.08 (0.19)	1.66 (0.33)	1.70 (0.46)	1.72 (0.29)	2.65 (0.21)	Pl > Sf = Pa	***	*
C:N	46.89 (8.92)	41.69 (6.96)	28.19 (5.11)	28.06 (7.18)	27.10 (4.27)	17.74 (1.48)	Pa = Sf > Pl	***	NS
δ <sup>15</sup> N	2.27 (1.55)	1.90 (0.97)	1.57 (1.34)	3.30 (0.80)	3.45 (0.66)	2.64 (0.98)	NS	***	NS
δ <sup>13</sup> C	-30.47 (0.91)	-30.79 (1.05)	-29.06 (1.30)	-29.34 (1.13)	-31.22 (1.01)	-30.96 (1.14)	Sf > Pa = Pl	NS	NS
R <sub>d</sub>	-0.62 (0.27)	-0.77 (0.59)	-0.54 (0.33)	-2.06 (1.31)	-0.44 (0.30)	-0.50 (0.22)	Pl = Sf > Pa	**	***
LCP	26 (19)	25 (28)	6 (3)	95 (27)	15 (14)	13 (16)	Pa > Sf = Pl	*	*
Φ	ND <sup>5</sup>	ND	ND	0.021 (0.011)	0.054 (0.071)	0.062 (0.049)	Pl ≥ Sf ≥ Pa	*	NS
PDF <sup>sat</sup>	404 (293)	314 (206)	320 (176)	375 (339)	291 (199)	356 (150)	NS	NS	NS
A <sub>max</sub>	3.14 (1.18)	3.49 (1.67)	4.46 (1.84)	4.15 (1.73)	2.71 (1.17)	5.35 (2.25)	Pl ≥ Pa ≥ Sf	NS	NS
F <sub>v</sub> /F <sub>M</sub>	0.456 (0.184)	0.615 (0.103)	0.698 (0.039)	0.378 (0.204)	0.653 (0.073)	0.692 (0.096)	Pl = Sf > Pa	NS	NS
ΦPSII <sup>700</sup>	0.086 (0.043)	0.071 (0.026)	0.115 (0.040)	0.109 (0.072)	0.084 (0.038)	0.142 (0.057)	Pl > Sf = Pa	*	NS
qp <sup>700</sup>	0.408 (0.098)	0.248 (0.064)	0.338 (0.110)	0.539 (0.157)	0.315 (0.133)	0.442 (0.113)	Pl = Pa > Sf	***	NS
D <sup>700</sup>	0.792 (0.082)	0.714 (0.064)	0.661 (0.031)	0.814 (0.098)	0.732 (0.047)	0.681 (0.072)	Pa > Sf > Pl	NS	NS
ETR <sup>700</sup>	25.6 (12.9)	21.2 (7.7)	34.2 (12.0)	32.4 (21.4)	24.9 (11.3)	42.4 (16.9)	Pl > Pa = Sf	*	NS
V <sub>cmax</sub>	ND	13.59 (1.76)	22.40 (3.70)	ND	11.18 (1.13)	20.77 (7.45)	Pl > Sf	NS	NS
J <sub>max</sub>	ND	12.06 (3.75)	21.18 (5.55)	ND	11.04 (1.71)	20.25 (10.15)	Pl > Sf	NS	NS
TPU	ND	2.06 (0.17)	3.00 (0.49)	ND	1.70 (0.13)	2.82 (0.77)	Pl > Sf	NS	NS

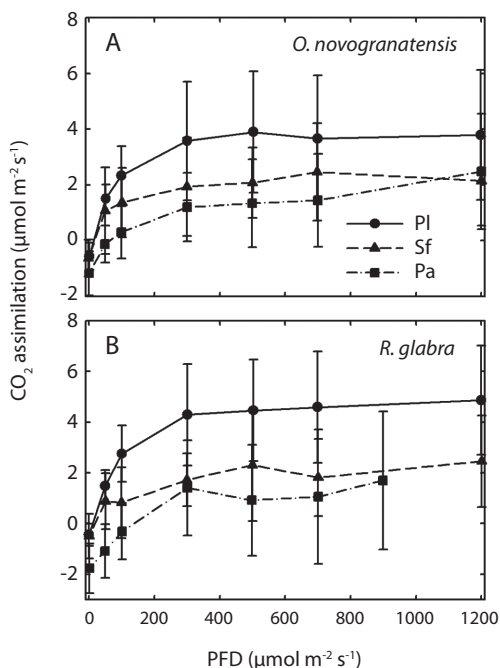
- For full statistical results see Appendix. Leaf area (cm<sup>2</sup>); SLA=specific leaf area (cm<sup>2</sup>/g); δ<sup>15</sup>N (‰); δ<sup>13</sup>C (‰); V<sub>cmax</sub>=carboxylation efficiency (mol/m<sup>2</sup>.s); J<sub>max</sub>=electron transport derived from A-C<sub>c</sub> curves (μmol/m<sup>2</sup>.s); TPU=triose phosphate utilization (μmol/m<sup>2</sup>.s); PFDsat=PFD at which light saturation occurs (μmol/m<sup>2</sup>.s); LCP=light compensation point (μmol/m<sup>2</sup>.s); Φ=apparent quantum yield of CO<sub>2</sub> uptake from A-Q curves (CO<sub>2</sub>/photon); A<sub>max</sub>=maximal light-saturated rate of CO<sub>2</sub> uptake from A-Q curves (μmol/m<sup>2</sup>.s); R<sub>d</sub>=dark respiration from A-Q curves (μmol/m<sup>2</sup>.s); F<sub>v</sub>/F<sub>M</sub>=maximal quantum efficiency of PSII (unitless); ΦPSII<sup>700</sup>=quantum efficiency of PSII measured at a PFD of 700 μmol/m<sup>2</sup>.s (unitless); qp<sup>700</sup>=the relative number of open PSII reaction centers measured at a PFD of 700 μmol/m<sup>2</sup>.s (unitless); D<sup>700</sup>=engagement of energy dissipation processes measured at a PFD of 700 μmol/m<sup>2</sup>.s (unitless); ETR<sup>700</sup>=electron transport rate through PSII (μmol/m<sup>2</sup>.s) measured at a PFD of 700 μmol/m<sup>2</sup>.s.
- Order of significant differences for the habitat comparisons. > indicates that a treatment or species had a significantly (p<0.05) higher value than another habitat using Tukey's LSD multiple comparison procedure. NS = not significant.
- Level of significance between species. Number of seedlings per species × habitat combination – n=17-21 for SLA & leaf area; n=10-15 for fluorescence and stable isotopes; n=4-15 for A-Q curves; n=4-5 for A-C<sub>c</sub> curves. \* = p<0.05, \*\* p<0.01, \*\*\* p<0.001.
- Level of significance for the habitat × species interaction term.
- “ND”: not determined due to lack of healthy leaf material.

light-saturated maximal rate of photosynthetic CO<sub>2</sub> uptake (A<sub>max</sub>) was higher in plantations than secondary forests, and was variable across species in pastures, likely due to the small sample size of surviving seedlings.

In general, PSII function decreased and thermal dissipation increased as a function of PFD for both species (Fig. 2). The apparent quantum efficiency of PSII for dark-adapted

leaves (F<sub>v</sub>/F<sub>M</sub>) was highest for seedlings in plantations and secondary forests, as compared to pastures (Table 3, Appendix). The quantum efficiency of PSII at a PFD of 700 μmol/m<sup>2</sup>.s (ΦPSII<sup>700</sup>) was highest in plantations. The proportion of open PSII reaction centers (qp<sup>700</sup>) was highest in pastures and lowest in secondary forests. Thermal dissipation (D<sup>700</sup>) was highest in pasture habitats. The electron transport rate





**Fig. 1.** Light response curves (“A-Q curves”) for *Otoba novogranatensis* and *Ruagea glabra*. Data are means $\pm$ SD for n=4-15 plants in Pasture (Pa), Secondary forest (Sf) and Plantation (Pl) habitats.

determined from PSII fluorescence ( $ETR^{700}$ ) was significantly higher in plantations and in *R. glabra*.

Based on the *A-Ci* curves (Fig. 3), both species had a higher photosynthetic capacity ( $V_{c_{max}}$ ,  $J_{max}$ , and TPU) in plantations than in secondary forests (Table 3, Appendix). Carboxylation efficiency ( $V_{c_{max}}$ ) was 1.9 (*R. glabra*) and 1.6 fold (*O. novogranatensis*) higher in plantations compared to secondary forests. Seedlings in the pasture had small and yellowed leaves that did not provide *A-Ci* curves that could be analyzed by the software.

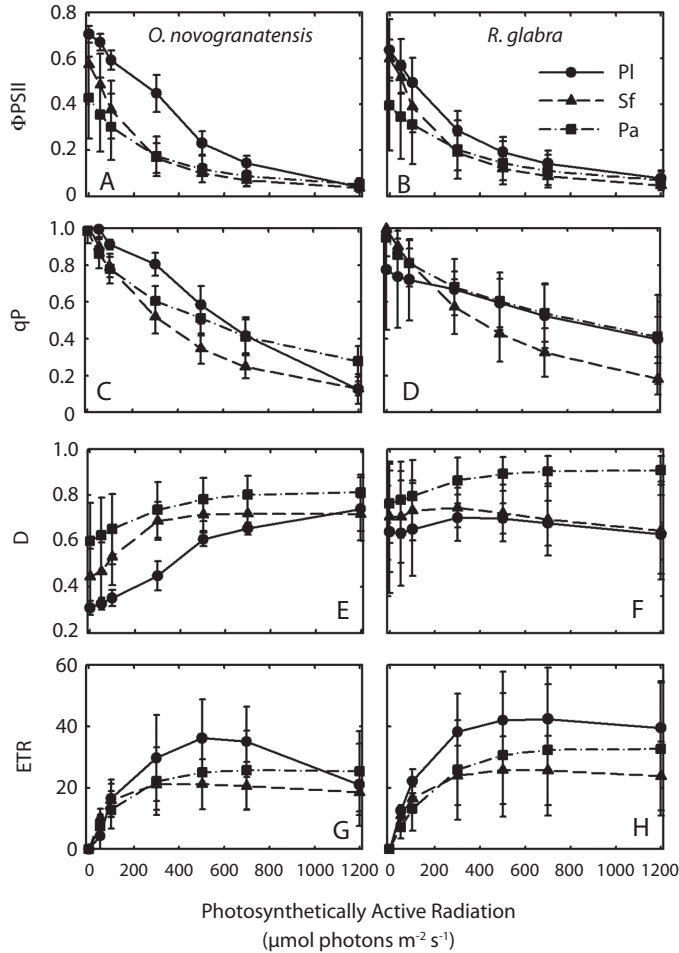
Water relations did not differ significantly across sites, species, or habitats. Mean volumetric soil moisture content was  $23.0\pm 5.7\%$  and ranged from 14.0 to 35.7% across all plots. Stem water potential varied from -0.15 to -1.80MPa and averaged  $-0.80\pm 0.47$  MPa. Carbon stable isotope ratios ( $\delta^{13}C$ ) showed a significant habitat  $\times$  species effect, and were highest (less negative) in plantations for *O.*

*novogranatensis*, yet highest in the pastures for *R. glabra* (Table 3).

## DISCUSSION

Past studies have suggested that enrichment planting is an effective strategy for introducing later successional species into degraded tropical forest ecosystems (Peña-Claros *et al.* 2002, Romell *et al.* 2008, Doucet *et al.* 2009). Prior research at our study sites showed that seedlings were taller, had lower root:shoot ratios, and higher leaf N content when they were planted in tree plantations, compared to secondary forests or abandoned pastures (Cole *et al.* 2011). Our current results show that in this moist forest ecosystem, planting seedlings in the same plantations (which contain some N-fixing tree species) can enhance certain aspects of photosynthetic gas exchange, biochemistry, and energy processing within PSII for seedlings of the two species we examined and that micrometeorological conditions may play a lesser role in explaining their growth differences.

Planting seeds under plantation trees resulted in seedlings with greater aboveground growth. The  $\delta^{15}N$  results suggest that some of the N in leaves of seedlings in plantations is from biological N-fixation by the trees, although we note that this requires further study given that we found marginally ( $p=0.0615$ ) lower  $\delta^{15}N$  in abandoned pastures. Higher leaf %N for seedlings in plantations is consistent with results showing more favorable biochemical characteristics associated with photosynthetic capacity and chlorophyll *a* fluorescence from Photosystem II (Bungard *et al.* 2002, Kitao *et al.* 2006, Cernusak *et al.* 2008, Reich *et al.* 2009). Moreover, the quantum efficiency of PSII in darkness ( $F_V/F_M$ ) and at  $700\mu\text{mol}/\text{m}^2\cdot\text{s}$  ( $\Phi_{PSII}^{700}$ ), the rate of electron transport ( $ETR^{700}$ ), and the degree of thermal dissipation ( $D^{700}$ ) are all consistent with greater leaf N in the plantation habitats. By contrast, leaves of seedlings in the pasture showed signs of chronic photoinhibition. It should be noted that previous research shows that not all species

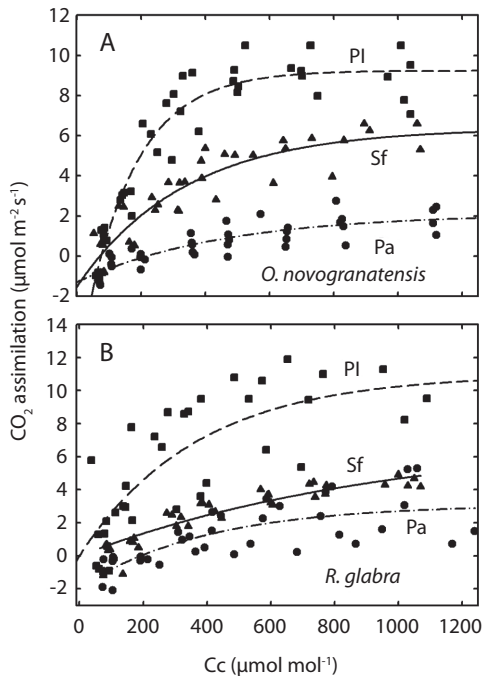


**Fig. 2.** Chlorophyll *a* fluorescence from PSII responses to PFD (400–700nm) for *Otoba novogranatensis* and *Ruagea glabra*. A, B. Quantum efficiency of PSII. C, D. The proportion of open reaction centers. E, F. The engagement of thermal dissipation processes. G, H. Electron transport rate. Data are means±SE for n=10-15 plants per habitat type. Unhealthy leaf tissue precluded measurements for *R. glabra* in pastures.

exhibit a causal relationship between leaf N and photosynthetic assimilation (Aragao *et al.* 2005), and leaf P and other nutrients affect carbon gain as well (Meir *et al.* 2001, Reich *et al.* 2009), which may have affected some of the growth differences observed by Cole *et al.* (2011).

For both species, in plantations there was greater carboxylation efficiency by activated Rubisco, electron transport to drive regeneration of the carboxylation substrate RuBP, and triose-phosphate utilization, as determined by

the *A-Ci* curves. These patterns are generally associated with an up-regulation of photosynthetic capacity in the presence of higher foliar N content (Coomes & Grubb 1998, Bungard *et al.* 2002, Coste *et al.* 2005, Reich *et al.* 2009), which could drive greater carbon gain, survival, and growth (Cole *et al.* 2011). Some aspects of the *A-Q* response (i.e. less-negative  $R_d$ , lower LCP, and higher  $A_{max}$ ) were consistent with higher photosynthetic capacity for seedlings from the tree plantations in comparison to pasture and secondary forest habitats



**Fig. 3.** CO<sub>2</sub> response curves (“A-Cc curves”) for *Oroba novogranatensis* and *Ruagea glabra* at the Finca Loma Linda site. Non-linear regressions were computed using SigmaPlot (SPSS, Inc. v 10) by combining data for all plants in Pasture (Pa), Secondary forest (Sf), and Plantation (Pl) habitats.

(Riddoch *et al.* 1991, Johnson *et al.* 1997, Ellis *et al.* 2000, Posada *et al.* 2009). When  $R_d$  and  $A_{max}$  are considered together, seedlings of both species in the pasture habitats had a lower net C budget compared to leaves from plants in plantations and secondary forests.

Higher N in seedlings in the plantations also appears to be associated with a greater flexibility to adjust PSII light harvesting. As expected with increasing light (PFD) levels,  $\Phi_{PSII}$  and  $qP$  decreased, and  $D$  and  $ETR$  increased. However, the magnitude of these effects differed across species, as determined by  $\Phi_{PSII}^{700}$ ,  $qP^{700}$  and  $ETR^{700}$ . The apparent quantum yield for chlorophyll fluorescence from PSII ( $F_v/F_M$ ) was significantly higher in the plantations and secondary forest habitats compared to the pastures. These species may

differ across habitats in terms of their ability to undergo flexible thermal energy dissipation via a trans-thylakoid  $\Delta pH$  and PsbS protein activity (Demmig-Adams & Adams 2006). The number of open PSII reaction centers was similar in the pastures and plantations, and lowest in the secondary forest habitats, which is more difficult to explain. Possibly the higher  $D$  in pastures allowed some PSII reaction centers to remain open, while higher leaf N in plantations accounted for higher  $ETR$ , with seedlings in secondary forests subject to lower leaf N and intermediate amounts of  $D$ . In other studies, some species show a down regulation of PSII efficiency under limited N (Verhoeven *et al.* 1997). It is possible that our results for  $\Phi_{PSII}$ ,  $ETR$  and  $D$  across habitats may be due to varying levels of foliar N content that can support production of violaxanthin de-epoxidase, PsbS protein, and other components of PSII (Li *et al.* 2002, Niyogi *et al.* 2005, Logan *et al.* 2008) beyond simply affecting activated Rubisco quantity.

Our results suggest that soil water content and micrometeorological factors may be of lesser importance than foliar N for driving differences in photosynthetic processes across planting habitats. Although overstorey cover was much higher and understorey cover lower in plantation and secondary forest habitats, micrometeorological conditions (PFD,  $T^{air}$ ,  $T^{soil}$  and RH) at seedling height were highly variable across sites and planting habitats, and did not show consistent patterns. The lack of micrometeorological differences across treatments at seedling height (~0.2m) can be explained partly by the vegetation structure. Seedlings were shaded by the tree canopy in plantation and secondary forest sites and by grass cover (1-1.5m height) in pastures, which has been shown to result in relatively similar microclimatic conditions (Holl 1999). Water deficit was also not an apparent factor in the survival and growth of seedlings across sites and treatments, given the lack of habitat differences in soil moisture, plant water potential, or integrated WUE over the lifetime of the leaf, as determined by  $\delta^{13}C$ .

In some cases, physiological responses of seedlings in the secondary forest habitats were similar to seedlings in pastures (e.g. for leaf area, %C, %N,  $\Phi$ PSII<sup>700</sup>, ETR<sup>700</sup>), whereas for other parameters responses in secondary forests were intermediate between those from pastures and plantations (e.g. SLA and D<sup>700</sup>), and in a few cases secondary forest values were similar to plantations (e.g. R<sub>d</sub>, LCP, F<sub>v</sub>/F<sub>M</sub>). The reason for these patterns is not clear, but we offer a few potential possibilities. First, previous work at this study site has shown that while leaf litter N content is higher in the plantations compared to pastures and secondary forests, annual leaf litterfall inputs of K, Ca, Mg and Zn are higher in secondary forests, likely due to the higher diversity of tree species (Celentano *et al.* 2011). Higher inputs of these other nutrients may counteract the effects of lower N for certain photosynthetic processes. Second, grass cover and bare ground in secondary forests were intermediate to plantations and pastures, so differential competition with understory species may have affected our seedling responses, although experiments directly manipulating understory competition would be necessary to test this hypothesis. Third, our single-point microclimatic measurements may not have adequately characterized the high spatial complexity of the physical conditions in the three habitats.

Overall, our results show that enrichment planting under a mixed-species canopy is a successful strategy for introducing these two later successional tree species as part of tropical forest restoration efforts. Our results suggest that N-fixing trees in plantations may enhance the photosynthetic capacity and PSII efficiency for seedlings of *O. novogranatensis* and *R. glabra* and perhaps other tropical forest tree species. But, the role of N-fixation by trees for facilitating seedlings in this system is still not clear. These results highlight the importance of the question of which species to include as part of tropical forest restoration plantings in order to maximize the return on the time and

finances invested to restore habitat diversity and functions.

## ACKNOWLEDGMENTS

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## RESUMEN

El enriquecimiento de bosques secundarios o plantaciones forestales en proceso de regeneración natural por medio de la siembra de plántulas es una práctica cada vez más utilizada para restaurar bosques tropicales en estado de sucesión tardía. Sembramos dos especies de árboles (*Otoba novogranatensis* y *Ruagea glabra*) en pastizales abandonados, bosques secundarios de ocho a diez años de edad y plantaciones forestales de tres años de edad (con dos especies fijadoras de Nitrógeno de un total de cuatro especies) en tres sitios agrícolas en el Sur de Costa Rica. Medimos condiciones micrometeorológicas, contenido de agua del suelo, potencial hídrico de las plantas, área foliar, C y N foliar, y fotosíntesis para entender de una mejor manera las respuestas funcionales de las plántulas ante condiciones de distintos estadios sucesionales. Las condiciones micrometeorológicas, contenido hídrico del suelo y el potencial hídrico de las plantas fueron mayoritariamente similares entre hábitats. Algunos aspectos de las hojas (como Área Foliar Específica y contenido de N foliar) y fotosíntesis (ej.: rendimiento cuántico y tasa de transporte de electrones) presentaron valores mayores en las plantaciones, intermedios en los bosques secundarios y menores en los pastizales abandonados. Se obtuvo un aumento en las tasas fotosintéticas bioquímicas (como  $V_{cmax}$ ,  $J_{max}$ ) y la eficiencia del Fotosistema II (ej.: disipación de energía térmica) en hojas provenientes de las plantaciones comparado a las de los pastizales, posiblemente relacionado a un mayor %N foliar. Los resultados sugieren que el N foliar puede ser más importante que el contenido de agua del suelo y que los factores micrometeorológicos para marcar diferencias en los procesos fotosintéticos entre hábitats. Las plántulas de estas dos especies en las plantaciones con árboles de tres años de edad (incluyendo dos fijadoras de N) incrementaron ciertos aspectos de su fotosíntesis y crecimiento comparado a las plántulas en los pastizales abandonados de especies exóticas, por lo tanto, esta práctica puede ayudar a incrementar la recuperación de los bosques en áreas agrícolas abandonadas.

**Palabras clave:** fluorescencia de clorofila a, restauración de bosques, intercambio gaseoso, contenido de N foliar, fotosistema II, bosque tropical secundario.

## REFERENCES

- Amiard, V., K.E. Mueh, B. Demmig-Adams, V. Ebbert, R. Turgeon & W.W. Adams. 2005. Anatomical and photosynthetic acclimation to the light environment in species with differing mechanisms of phloem loading. *Proc. Nat. Acad. Sci. USA* 102: 12968-12973.
- Anderson, J.M., W.S. Chow & J. De Las Rivas. 2008. Dynamic flexibility in the structure and function of photosystem II in higher plant thylakoid membranes: the grana enigma. *Photosynth. Res.* 98: 575-587.
- Aragao, D.V., L.B. Fortini, S. Mulkey, D.J. Zarin, M.M. Araujo & C.J.R. De Carvalho. 2005. Correlation but no causation between leaf nitrogen and maximum assimilation: The role of drought and reproduction in gas exchange in an understory tropical plant *Miconia ciliata* (Melastomataceae). *Am. J. Bot.* 92: 456-461.
- Bonilla-Moheno, M. & K.D. Holl. 2010. Direct seeding to restore tropical mature-forest species in areas of slash-and-burn agriculture. *Restor. Ecol.* 18: 438-445.
- Bungard, R.A., S.A. Zipperlen, M.C. Press & J.D. Scholes. 2002. The influence of nutrients on growth and photosynthesis of seedlings of two rainforest dipterocarp species. *Funct. Plant Biol.* 29: 505-515.
- Camargo, J.L.C., I.D.K. Ferraz & A.M. Imakawa. 2002. Rehabilitation of degraded areas of central Amazonia using direct sowing of forest tree seeds. *Restor. Ecol.* 10: 636-644.
- Celentano, D., R.A. Zahawi, B. Finegan, R. Ostertag, R.J. Cole & K.D. Holl. 2011. Litterfall dynamics under different tropical forest restoration strategies. *Biotropica* 43: 279-287.
- Cernusak, L.A., K. Winter, J. Aranda & B.L. Turner. 2008. Conifers, angiosperm trees, and lianas: Growth, whole-plant water and nitrogen use efficiency, and stable isotope composition ( $\delta C-13$  and  $\delta O-18$ ) of seedlings grown in a tropical environment. *Plant Physiol.* 148: 642-659.
- Chazdon, R.L. 2008. Beyond deforestation: Restoring forests and ecosystem services on degraded lands. *Science* 320: 1458-1460.
- Cole, R.J., K.D. Holl & R.A. Zahawi. 2010. Seed rain under tree islands planted to restore degraded lands in a tropical agricultural landscape. *Ecol. App.* 20: 1255-1269.
- Cole, R.J., C. Keene, R.A. Zahawi & K.D. Holl. 2011. Direct seeding of late successional trees to restore tropical montane forest. *For. Ecol. Manage.* 261: 1590-1597.
- Coomes, D.A. & P.J. Grubb. 1998. Responses of juvenile trees to above- and belowground competition in nutrient-starved Amazonian rain forest. *Ecology* 79: 768-782.
- Coste, S., J.C. Roggy, P. Imbert, C. Born, D. Bonal & E. Dreyer. 2005. Leaf photosynthetic traits of 14 tropical rain forest species in relation to leaf nitrogen concentration and shade tolerance. *Tree Physiol.* 25: 1127-1137.
- Cusack, D. & F. Montagnini. 2004. The role of native species plantations in recovery of understory woody diversity in degraded pasturelands of Costa Rica. *For. Ecol. Manage.* 188: 1-15.
- Demmig-Adams, B. & W.W. Adams. 2006. Photoprotection in an ecological context: the remarkable complexity of thermal energy dissipation. *New Phytol.* 172: 11-21.
- Doucet, J.L., Y.L. Kouadio, D. Monticelli & P. Lejeune. 2009. Enrichment of logging gaps with moabi (*Bailonella toxisperma* Pierre) in a Central African rain forest. *For. Ecol. Manage.* 258: 2407-2415.
- Ellis, A.R., S.P. Hubbell & C. Potvin. 2000. In situ field measurements of photosynthetic rates of tropical tree species: a test of the functional group hypothesis. *Can. J. Bot.* 78: 1336-1347.
- Engel, V.L. & J.A. Parrotta. 2001. An evaluation of direct seeding for reforestation of degraded lands in central São Paulo state, Brazil. *For. Ecol. Manage.* 152: 169-181.
- Holdridge, L.R., W.C. Grenke, W.H. Hatheway, T. Liang & J.A. Tosi Jr. 1971. Forest environments in tropical life zones. Pergamon, Oxford.
- Holl, K.D. 1999. Factors limiting tropical rain forest regeneration in abandoned pasture: seed rain, seed germination, microclimate, and soil. *Biotropica* 31: 229-241.
- Holl, K.D. 2002. Tropical moist forest, p. 539-558. *In* M.R. Perrow & A.J. Davy (eds.). Handbook of ecological restoration. Cambridge University, Cambridge, United Kingdom.
- Holl, K.D. & T.M. Aide. 2011. When and where to actively restore ecosystems? *For. Ecol. Manage.* 261: 1588-1563.
- Holl, K.D., R.A. Zahawi, R.J. Cole, R. Ostertag & S. Cordell. 2011. Planting seedlings in plantations versus tree islands as a large-scale tropical forest restoration strategy. *Restor. Ecol.* 19: 470-479.
- Johnson, J.D., R. Tognetti, M. Michelozzi, S. Pinzauti, G. Minotta & M. Borghetti. 1997. Ecophysiological responses of *Fagus sylvatica* seedlings to changing light conditions. The interaction of light environment and soil fertility on seedling physiology. *Physiol. Plantarum* 101: 124-134.

- Kitao, M., R. Yoneda, H. Tobita, Y. Matsumoto, Y. Maruyama, A. Arifin, A.M. Azani & M.N. Muhamad. 2006. Susceptibility to photoinhibition in seedlings of six tropical fruit tree species native to Malaysia following transplantation to a degraded land. *Trees-Struct. Funct.* 20: 601-610.
- Lamb, D., P.D. Erskine & J.D. Parrotta. 2005. Restoration of degraded tropical forest landscapes. *Science* 310: 1628-1632.
- Li, X.P., P. Muller-Moule, A.M. Gilmore & K.K. Niyogi. 2002. PsbS-dependent enhancement of feedback de-excitation protects Photosystem II from photoinhibition. *Proc. Nat. Acad. Sci. USA* 99: 15222-15227.
- Logan, B.A., S.G. Terry & K.K. Niyogi. 2008. Arabidopsis genotypes with differing levels of psbS expression differ in Photosystem II quantum yield, xanthophyll cycle pool size, and aboveground growth. *Int. J. Plant Sci.* 169: 597-604.
- Loik, M.E. & K.D. Holl. 1999. Photosynthetic responses to light for rainforest seedlings planted in abandoned pasture, Costa Rica. *Restor. Ecol.* 7: 382-391.
- Long, S.P. & C.J. Bernacchi. 2003. Gas exchange measurements, what can they tell us about the underlying limitations to photosynthesis? Procedures and sources of error. *J. Exp. Bot.* 54: 2393-2401.
- Martínez-Garza, C. & H.F. Howe. 2003. Restoring tropical diversity: beating the time tax on species loss. *J. App. Ecol.* 40: 423-429.
- Meir, P., J. Grace & A.C. Miranda. 2001. Leaf respiration in two tropical rainforests: constraints on physiology by phosphorus, nitrogen and temperature. *Funct. Ecol.* 15: 378-387.
- Minchin, P.E.H. & A. Lacoite. 2005. New understanding on phloem physiology and possible consequences for modelling long-distance carbon transport. *New Phytol.* 166: 771-779.
- Niyogi, K.K., X.P. Li, V. Rosenberg & H.S. Jung. 2005. Is PsbS the site of non-photochemical quenching in photosynthesis? *J. Exp. Bot.* 56: 375-382.
- Pammenter, N.W., F. Loreto & T.D. Sharkey. 1993. End-product feedback effects on photosynthetic electron-transport. *Photosynth. Res.* 35: 5-14.
- Peña-Claros, M., R.G.A. Boot, J. Dorado-Lora & A. Zonta. 2002. Enrichment planting of *Bertholletia excelsa* in secondary forest in the Bolivian Amazon: effect of cutting line width on survival, growth and crown traits. *For. Ecol. Manage.* 161: 159-168.
- Piotto, D. 2007. Growth of native tree species planted in open pasture, young secondary forest and mature forest in humid tropical Costa Rica. *J. Trop. For. Sci.* 19: 92-102.
- Pons, T.L. & R.A.M. Welschen. 2002. Overestimation of respiration rates in commercially available clamp-on leaf chambers. Complications with measurement of net photosynthesis. *Plant Cell Environ.* 25: 1367-1372.
- Posada, J.M., M.J. Lechowicz & K. Kitajima. 2009. Optimal photosynthetic use of light by tropical tree crowns achieved by adjustment of individual leaf angles and nitrogen content. *Ann. Bot-London* 103: 795-805.
- Raines, C.A. 2003. The Calvin cycle revisited. *Photosynth. Res.* 75: 1-10.
- Reich, P.B., J. Oleksyn & I.J. Wright. 2009. Leaf phosphorus influences the photosynthesis-nitrogen relation: a cross-biome analysis of 314 species. *Oecologia* 160: 207-212.
- Riddoch, I., T. Lehto & J. Grace. 1991. Photosynthesis of tropical tree seedlings in relation to light and nutrient supply. *New Phytol.* 119: 137-147.
- Romell, E., G. Hallsby, A. Karlsson & C. García. 2008. Artificial canopy gaps in a *Macaranga* spp. dominated secondary tropical rain forest - Effects on survival and above ground increment of four underplanted dipterocarp species. *For. Ecol. Manage.* 255: 1452-1460.
- Schatral, A. & J.E.D. Fox. 1994. Quality and viability of seeds in the genus *Hibbertia*. *Seed Sci. Technol.* 22: 273-284.
- Siddique, I., V.L. Engel, J.A. Parrotta, D. Lamb, G.B. Nardoto, J. Ometto, L.A. Martinelli & S. Schmidt. 2008. Dominance of legume trees alters nutrient relations in mixed species forest restoration plantings within seven years. *Biogeochemistry* 88: 89-101.
- Sims, D.A. & R.W. Pearcy. 1991. Photosynthesis and respiration in *Alocasia-macrorrhiza* following transfers to high and low light. *Oecologia* 86: 447-453.
- Vankooten, O. & J.F.H. Snel. 1990. The use of chlorophyll fluorescence nomenclature in plant stress physiology. *Photosynth. Res.* 25: 147-150.
- Verhoeven, A.S., B. DemmigAdams & W.W. Adams. 1997. Enhanced employment of the xanthophyll cycle and thermal energy dissipation in spinach exposed to high light and N stress. *Plant Physiol.* 113: 817-824.
- Zimmerman, J.K., T.M. Aide & A.E. Lugo. 2007. Implications of land use history for natural forest regeneration and restoration strategies in Puerto Rico, p. 51-74. *In* R.J. Hobbs & V.A. Cramer (eds.). *Old fields*, Island, Washington, DC, USA.

APPENDIX  
Summary of statistical results from two way analyses of variance

Parameter	Site	Habitat	Species	Hab × Spec	Df err
Leaf area	2.2 (0.0084)	89.3 (<0.0001)	22.9 (<0.0001)	9.3 (0.0002)	113
SLA	2.4 (0.0741)	17.3 (<0.0001)	91.1 (<0.0001)	3.1 (0.0503)	113
%C	8.8 (0.0004)	14.5 (<0.0001)	30.4 (<0.0001)	1.9 (0.1524)	72
%N	3.4 (0.0382)	76.6 (<0.0001)	149.4 (<0.0001)	3.6 (0.0326)	72
C:N	4.3 (0.0180)	50.0 (<0.0001)	118.2 (<0.0001)	2.6 (0.0799)	72
$\delta^{15}\text{N}$	5.7 (0.0051)	2.8 (0.0681)	24.2 (<0.0081)	0.9 (0.4292)	70
$\delta^{13}\text{C}$	0.23 (0.7953)	8.0 (0.0007)	2.5 (0.1205)	11.6 (<0.0001)	72
PDF <sup>sat</sup>	1.8 (0.1807)	0.4 (0.6941)	0.1 (0.8107)	0.3 (0.7501)	58
LCP	2.2 (0.1217)	15.3 (<0.0001)	5.1 (0.0284)	4.2 (0.0207)	58
$\Phi$	1.2 (0.3181)	6.2 (0.0035)	4.9 (0.0315)	0.1 (0.9576)	58
$A_{\text{max}}$	0.1 (0.9170)	7.1 (0.0017)	0.5 (0.4825)	1.7 (0.1972)	58
$R_{\text{d}}$	0.1 (0.9465)	11.7 (<0.0001)	7.4 (0.0088)	12.5 (<0.0001)	58
$F_{\text{v}}/F_{\text{M}}$	13.9 (<0.0001)	41.4 (<0.0001)	0.2 (0.6223)	1.5 (0.2356)	77
$\Phi\text{PSII}^{700}$	0.8 (0.4611)	9.2 (0.0003)	4.5 (0.0382)	0.2 (0.8110)	77
qp <sup>700</sup>	1.3 (0.2668)	20.5 (<0.0001)	17.4 (<0.0001)	0.7 (0.4933)	77
D <sup>700</sup>	12.8 (<0.0001)	33.0 (<0.0001)	2.4 (0.1262)	0.01 (0.9892)	77
ETR <sup>700</sup>	0.8 (0.4615)	9.2 (0.0003)	4.5 (0.0378)	0.2 (0.8076)	77
$V_{\text{cmax}}$	NA <sup>2</sup>	25.1 (0.0002)	0.2 (0.2885)	0.1 (0.8326)	15
$J_{\text{max}}$	NA	11.9 (0.0036)	0.1 (0.7286)	0.0 (0.9879)	15
TPU	NA	25.5 (0.0001)	1.8 (0.2027)	0.2 (0.6670)	15

1. Values are F (p) for site blocking term, habitat, species, and habitat × species interaction, and the degrees of freedom.
2. NA= Not applicable.

