

TWO NEW SPECIES OF *ECHINOSEPALA* (ORCHIDACEAE: PLEUROTHALLIDINAE)

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ABSTRACT. We describe and illustrate two new species of *Echinosepala* from Costa Rica, they are placed within a phylogenetic framework of the genus, and compared to their closest relatives. A short resume of the genus' history is given, reviewing the main characters that have been used to group its species at various taxonomic ranks. *Echinosepala expolita* is compared with *E. pan*, from which it mainly differs by the glabrous, inflated sheaths that cover the ramicaul, the distinctly broader ligulate-lanceolate leaves, the lanate-hirsute ovary, the much larger flower, and the tomentose abaxial indumentum of the sepals. *Echinosepala longipedunculata* is similar to *E. aspasicensis*, but can be recognized by the unusually long peduncle, the greenish yellow flower, the verrucose-tuberculate adaxial surface of the sepals, and the shorter, truncate, orange lip. A key to the Costa Rican species of *Echinosepala* is provided. A new combination in *Echinosepala* is proposed for *Acianthera biseta* from Peru.

RESUMEN. Se describen e ilustran dos nuevas especies de *Echinosepala* de Costa Rica, discutiéndolas dentro del marco filogenético del género y comparándolas con sus parientes más cercanos. Retomamos brevemente la historia del género, revisando los principales caracteres que han sido utilizados para agrupar sus especies en diferentes rangos taxonómicos. *Echinosepala expolita* se compara con *E. pan*, de la cual difiere principalmente por las vainas glabras e infladas que cubren el ramicaule, las hojas distintamente ligulado-lanceoladas, el ovario lanado-hirsuto, la flor mucho más grande y la indumentaria abaxial tomentosa de los sépalos. *Echinosepala longipedunculata* es similar a *E. aspasicensis*, pero puede ser reconocida por el pedúnculo inusualmente largo, las flores amarillo-verdosas, la superficie adaxial verrugosa-tuberculosa de los sépalos y el labelo más corto, truncado y de color naranja. Se proporciona una clave taxonómica para las especies costarricenses de *Echinosepala*. Se propone una nueva combinación en *Echinosepala* para *Acianthera biseta*.

KEY WORDS: *Acianthera*, *Echinosepala aspasicensis*, *Echinosepala pan*, flora of Costa Rica, *Myoxanthus*, new species, phylogenetics, pseudo-glenion

Introduction. Among the groups of “basal” Pleurothallidinae (*sensu* Karremans 2016), few other genera have been so difficult to place into a correct phylogenetic context as *Echinosepala* Pridgeon & M.W.Chase. Of the twelve to fourteen species belonging to the genus (IPNI 2017, Epidendra 2017), none were originally described in *Echinosepala*: one was proposed in *Acianthera* Scheidw., two in *Myoxanthus* Poepp. & Endl., and 11 in *Pleurothallis* R.Br. When Carlyle Luer (1992) first recognized that most species of the group were closely related and gathered them

into *Myoxanthus* subgen. *Silenia* Luer (typified by *Pleurothallis aspasicensis* Rchb.f.) and *Myoxanthus* subgen. *Satyria* Luer (typified by *Pleurothallis pan* Luer), all the species of *Echinosepala* but five had already been described as members of *Pleurothallis*. Luer (1992) distinguished subgen. *Satyria* and subgen. *Silenia* from other groups of *Myoxanthus* by the non-scurfy sheaths [except *Myoxanthus pan* (Luer) Luer \equiv *Echinosepala pan* (Luer) Pridgeon & M.W.Chase], and the flowers produced from low on the ramicaul or terminally at the base of the leaf (instead of from near

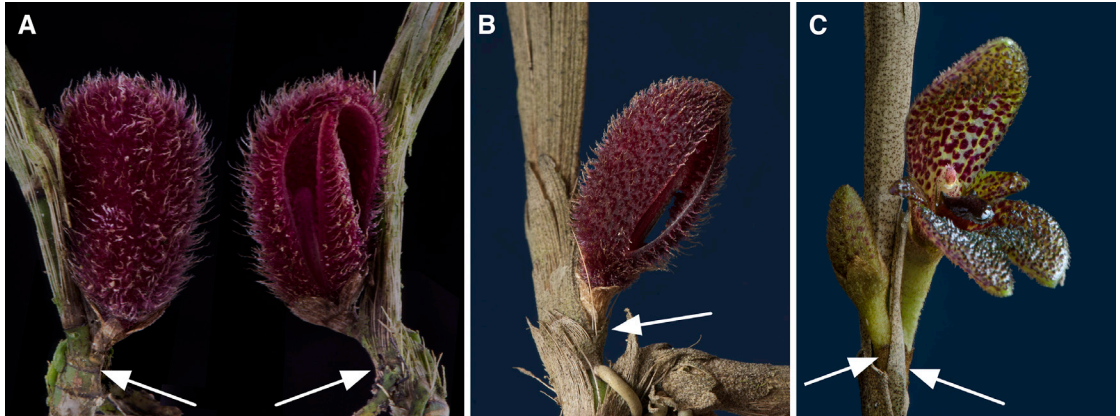


FIGURE 1. Basal inflorescences in species of *Echinosepala*. Note that the peduncle is exerted from a node of the stem (arrows). A, *E. lappiformis* (Bogarín 890). B, *E. stonei* (Bogarín 1871). C, *E. expolita* (Bogarín 1871). All the vouchers at JBL. Photographs by F. Pupulin.

the apex of the stem). The conspicuously verrucose-ciliate upper margin of the anther cap, although not unique to these species, is a common synapomorphy of the two subgenera. Considering that the species of *Myoxanthus* subgenera *Satyria* and *Silenia* had greater affinities with *Pleurothallis* subgenus *Acianthera* (Scheidw.) Luer, Luer (1995) moved most species back to *Pleurothallis*, demoting the two subgenera to sections within the subgen. *Acianthera* (Luer 1995, validated in Luer 1996). Before recombining some leftover species in *Echinosepala* (Luer 2007), Luer also suggested an affinity of the genus with *Kraenzlinella* Kuntze, transferring to it his *Pleurothallis shuarii* (Luer 1982, 2004). The analyses of DNA data (Pridgeon, Solano & Chase 2001, Pridgeon & Chase 2001) confirmed that the species of *Myoxanthus* subgen. *Silenia* and subgen. *Satyria* form a monophyletic group deserving generic recognition, and Pridgeon and Chase (2001) gave it formal status by creating *Echinella* Pridgeon & M.W.Chase [later corrected to *Echinosepala* Pridgeon & M.W.Chase (2002)].

The phylogenetic analyses also confirmed a close relationship between *Echinosepala* and *Myoxanthus*, as originally noted by Luer on the basis of morphological similarities. These two genera, together with *Barbosella* Schltr., *Dresslerella* Luer, *Pleurothallopsis* Porto & Brade, *Restrepia* Kunth, and *Restrepiella* Garay & Dunst., form one of the nine major affinities within the Pleurothallidinae, sister to the “basal” clade of *Octomeria* R.Br., and consecutively sister to the *Acianthera* clade.

Morphologically, the species of *Echinosepala*

are characterized by caespitose to repent plants with monophyllous ramicauls covered by tubular-ancipitous, minutely warty to pubescent sheaths; the leaf coriaceous; the inflorescence solitary-flowered, produced from the apical or lower nodes of the stem; the flower fleshy, with the sepals frequently hirsute abaxially and variously verrucose adaxially, the lateral sepals connate into a concave to cymbiform synsepal; the lip 3-lobed, with a high basal callus and with erect lateral lobes oblong to uncinately; the anther apical, incumbent, the anther cap papillose to ciliate at the apex; pollinia 2, with whale-tail shaped caudiculae (Luer 1992, Pridgeon 2005a).

The point of insertion of the inflorescence along the stem has been used as the main character to distinguish two assemblages of species within the group. Luer (1992) originally created subgen. *Satyria* of *Myoxanthus* to separate those species with a basal inflorescence, erroneously reported as produced from the rhizome (e.g., Luer 1992, Dressler 1993, Luer 2003, Pridgeon 2005a), from the rest of the group, which supposedly has a terminal inflorescence. Actually, the inflorescences of *Echinosepala* species are never rhizomic, and it is dubious that flowering from the nodes of the rhizome occurs at all in the Pleurothallidinae. The stem (ramicaul) of *Echinosepala* species is composed of 4 to 5 internodes which are completely enfolded by large, compressed-ancipitous, dry-papyraceous sheaths, the upper ones somewhat looser. Once the sheaths are removed, it is easy to observe that the basal inflorescence is produced along the stem, where it is exerted from the apex of the second to third internode (Fig. 1). The

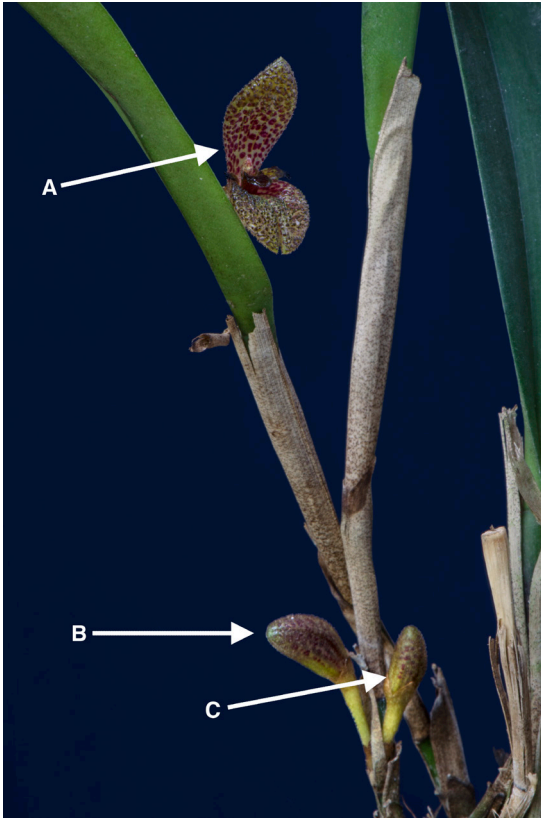


FIGURE 2. Simultaneous apical and basal inflorescences of *E. expolita* (arrows) (Pupulin 7030, JBL). Photograph by F. Pupulin.

production of the inflorescence from the lower nodes of the stem is apparently an obligate feature in *E. lappiformis* (A.H.Heller & L.O.Williams) Pridgeon & M.W.Chase, *E. pan*, and *E. stonei* (Luer) Pridgeon & M.W.Chase, but it is facultative in the species hereafter described as *E. expolita*, where both apical and basal inflorescences can be produced at once (Fig. 2). This condition has evolved in other groups of Pleurothallids, like *Acianthera*, and even in *Echinosepala* it could have arisen independently in two different groups of species.

The “basal” position of the inflorescence in species of *Echinosepala* convinced Luer (2004) to revive the genus *Brenesia* Schlechter (1923), originally coined to accommodate the anomalous *Brenesia costaricensis* Schltr., a species with a pendent inflorescence produced from the lower nodes of the ramicaul bearing hirsute flowers, and to transfer to *Brenesia* eight species previously treated under *Echinosepala* (Luer 2004), as well as *Pleurothallis johnsonii* Ames [\equiv *Acianthera*

johnsonii (Ames) Pridgeon & M.W.Chase], in the belief that they were related to each other. However, the relationship between species of *Brenesia stricto sensu* [i.e., *B. costaricensis*, *B. herrerae* (Luer) Luer, *B. johnsonii* (Ames) Luer + *Acianthera sotoana* Solano] and species of *Echinosepala* was not supported by the DNA analysis carried out by Pridgeon and collaborators (2001). More recently, a broad phylogenetic analysis of *Acianthera* (Karremans *et al.* 2016) showed that the clade including the type species of *Brenesia* is sister to a clade that includes *Antilla* (Luer) Luer and *Kraenzlinella*, altogether highly supported as sisters to the rest of *Acianthera*. The four Central American species of true “*Brenesia*” were formally recognized as a group with subgeneric status as *Acianthera* subgen. *Brenesia* (Schltr.) Karremans (Karremans *et al.* 2016).

Florally, species of *Echinosepala* are mostly characterized by the putrid color of the bilabiate perianth, often spreading only partially, and the complicated abaxial indumenta of the sepals. The indumentum varies from sparsely warty-verruculose [*E. aspasicensis*, *E. sempergemmata* (Luer) Pridgeon & M.W.Chase, *E. uncinata* (Fawc.) Pridgeon & M.W.Chase], the warts sometimes grouped into small tufts [*E. longipedunculata*, hereafter, and *E. vittata* (Pupulin & M.A.Blanco) C.O.Morales & N.Villalobos]; to sparsely spiny (*E. expolita*, hereafter); densely tomentose [*E. tomentosa* (Luer) Pridgeon & M.W.Chase]; hirsute with soft trichomes [*E. lappiformis* (A.H.Heller & L.O.Williams) Pridgeon & M.W.Chase, *E. stonei* (Luer) Pridgeon & M.W.Chase]; or echinate with stiff hairs (*E. pan*). Adaxially, the sepals are almost glabrous (*E. aspasicensis*, *E. sempergemmata*, *E. uncinata*, *E. vittata*); sparsely warty (*E. tomentosa*); to covered with irregular, low (*E. lappiformis*) or high (*E. stonei*) verrucae, or stiff, short, rounded (*E. longipedunculata*) or pointed (*E. expolita*, *E. pan*) warts. In two species the base of the dorsal sepal is semitransparent (*E. aspasicensis*, *E. expolita*), forming a window under the dark purple blotches (Fig. 3). The petals are mostly glabrous to sparsely verruculose, but they may be distinctly warty toward the apex (*E. expolita*, *E. lappiformis*, *E. pan*, *E. stonei*). The lip is glabrous in most species, but it is warty at the apex in *E. sempergemmata* and *E. shuarii* (Luer) Luer. At the base, it always presents a high callus which can be rounded and broad, massive, slightly concave, smooth or transversely rugose, occupying the entire



FIGURE 3. Fenestrate dorsal sepal in *E. aspicensis* (A, Karremans 4407) and *E. expolita* (B, Bogarín 1871). Vouchers at JBL. Photographs by F. Pupulin.

lip base (*E. lappiformis*, *E. sempergemmata*, *E. stonei*, *E. uncinata*), or provided with a horseshoe-shaped, concave pseudo-glenion (*E. aspicensis*, *E. expolita*, *E. longipedunculata*, *E. pan*, *E. tomentosa*, *E. vittata*). While true glenions in the Pleurothallidinae are rounded to elliptic to horseshoe-shaped depressions, often on the top of a raised area at the base of the lip, which apparently do not produce any exudates, the structure here defined as a pseudo-glenion is distinctly raised above the surface of the basal callus, and is provided with definite, apically rounded walls. Inside the walls a gelatinous, non-viscous, shiny exudate is produced, which is spread through a longitudinal groove to the apex of the lip, where it apparently dries, becoming matte, in two to three days (Fig. 4). In most species, there is a pair of intramarginal low lamellae, typically undulate on the upper margin.

Taxonomy. The first species belonging to *Echinosepala* was described in 1855 as *Pleurothallis aspicensis* from Colombia (Reichenbach 1855), followed by *Pleurothallis uncinata* Fawcett (1895), from a Jamaican collection. Both species have been treated as taxa with broad geographic distribution: *E. aspicensis* ranging from Costa Rica to Bolivia, the Guyanas, and Brazil, and *E. uncinata* recorded from the West Indies and Central America (Luer 1992). The correct application of both names is nonetheless problematic.

The type specimen of *E. aspicensis* at Kew (*Wagener 618!*) just consists of a crumpled flower from which the lip is missing, and no vegetative material has been conserved. Reichenbach (1855) described the lip as ligulate, attenuate at the retuse apex, provided with two small falcate lobes at the base, with a transverse keel on the disc and two small



FIGURE 4. Labella of *Echinosepala* species. On the left of the dashed line (A–K), labella with pseudo-glenion. On the right of dashed line (L–S), species without pseudo-glenion. A–C, *E. aspasicensis* (Bogarin 1945, JBL–27660, JBL–02078, respectively). D, *E. expolita* (Bogarin 1871). E, *E. pan* (Bogarin 10274). F–H, *E. longipedunculata* (Bogarin 3863, Bogarin 5449, Karremans 4494, respectively). I, *E. stonei* (Bogarin 7191). K, *E. tomentosa* (Bogarin 5622). L–N, *E. sempergemmata* (Bogarin 4601, Karremans 3199, Bogarin 4678, respectively). P–S, *E. uncinata* (Bogarin 5855, Bogarin 5871, Whitten 2153, respectively). Scale bar = 1 cm. All the vouchers at JBL. Photographs by F. Pupulin.

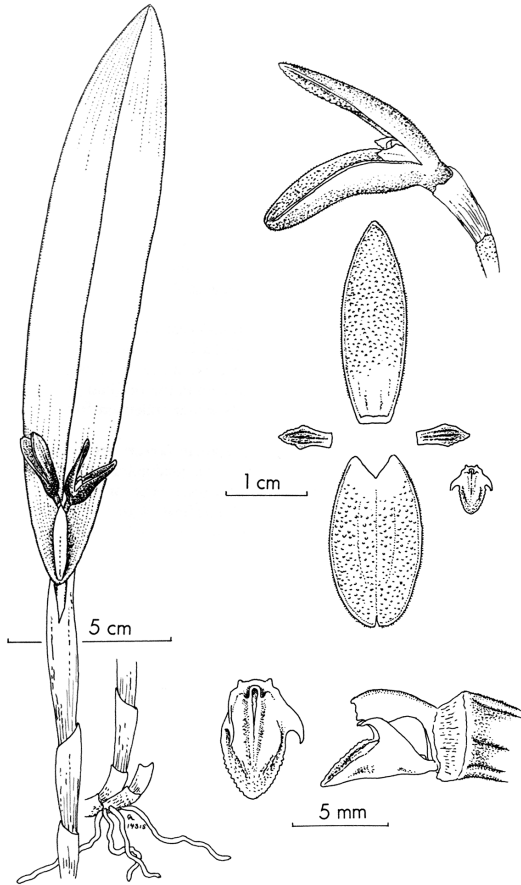


FIGURE 5. A Colombian specimen of *Echinosepala* (as *Myoxanthus*) *aspasicensis* (Luer 14315), from Luer 1992 (pl. 45). Courtesy of the Missouri Botanical Garden.

keels in front of the lateral lobes, running parallel to the margins of the midlobe; he did not mention a horseshoe-shaped, concave structure sitting on the basal callus that we refer to in this paper as pseudoglenion. The only modern documentation of a Colombian plant that we are aware of is that illustrated by Luer (1992: pl. 45), but this specimen was collected some 200 kilometers south of the type locality, along the western slopes of the Eastern Cordillera. Luer's specimen shows a plant with two bilabiate flowers, with a short, broad, elliptic lip (Fig. 5), quite distinct from Costa Rican populations treated under *E. aspasicensis* (Luer 2003 [under *Pleurothallis*], Pridgeon 2005b), which typically present a single flower and a narrow, elliptic-oblong, elongate lip. Only an effort to document the morphological and

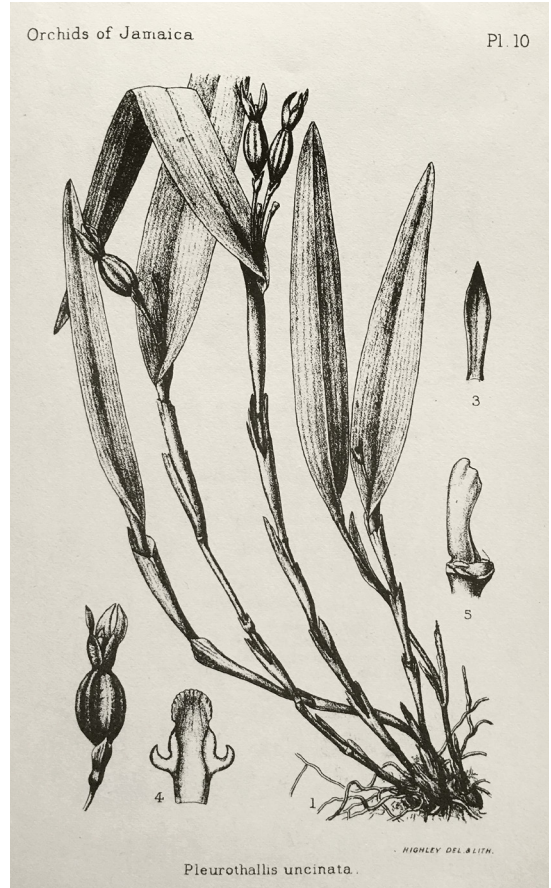


FIGURE 6. *Echinosepala* (as *Pleurothallis*) *uncinata* from Jamaica, from Fawcett & Rendle 1910 (pl. 10).

genetic variation of *Echinosepala* species at the type locality, near Aspasica in Colombia, could resolve the identity of the true *E. aspasicensis*, allowing the proper identification of similar taxa across the alleged distribution of the species.

The type specimen of *E. uncinata* (Harris s.n., K 584080!), on the other hand, shows a large plant, with ramicauls almost 20 cm long, which contrast with populations from Costa Rica that usually have ramicauls only 10–14 cm in length, and in size approaches instead Costa Rican specimens of *E. sempergemmata*. The floral parts of the type specimen consist of a floral bud and a fruit, and also the drawing of the species prepared by Percy Highley (1856–1929) for Fawcett and Rendle's treatment of Orchidaceae for *Flora of Jamaica* (Fawcett & Rendle 1910) depicts a

large plant with only pollinated flowers (Fig. 6). These are described as “scabrous” (Fawcett 1895) or “rough on inner and outer sides, tomentose on outer side” (Fawcett & Rendle 1910), a character that contrasts with Costa Rican populations treated under *E. uncinata* (which have glabrous flowers), and agrees instead with *E. sempergemmata*, with which we strongly suspect it is conspecific.

In preparation of a systematic treatment of *Echinosepala* for the flora of Costa Rica, we confidently describe in this paper two new species in the genus, as they cannot be confused with any of the previously published names. The Costa Rican *Pleurothallis alexandrae* Schltr., which is obviously a species of *Echinosepala*, belongs to the group of species that present a broad and smooth callus at the base of the lip, while both the new taxa proposed here have a basal pseudo-glenion. Of the new taxa, one is close to *E. pan*, a species that otherwise has no close relatives. The other is close to *A. aspasicensis sensu lato*, but it has a very long peduncle and a truncate lip that were not previously recorded in any other species of the genus.

Materials and methods. Plants used for this study were collected around the country and brought to Lankester Botanical Garden (JBL) for cultivation and subsequent documentation. Whenever possible, type localities and critical localities from where rare and/or unique species records were previously reported were visited for plant collecting and subsequent study. At least five specimens per morphospecies were collected at any given locality whenever possible. Field notes were taken during collections, including GPS and political data, elevation, ecological zones and main types of vegetation. Plants were cultivated at JBL, recording phenological data.

Morphological analysis.— We studied and documented over 150 individuals within this group, belonging to virtually all the morphospecies known for the country (Pupulin *et al.*, in prep.). Notes on floral ecology were taken through direct observation of the plants under the semi-artificial conditions of open greenhouses. Several plants in this group were documented at flowering with high-resolution images of the flower provided with a scale, often including

views of the whole plant and floral details with relative scales. Photographs were taken with Nikon cameras (D5200, D7100, D810) fitted with macro lenses (Micro Nikkor 60mm f/2.8, Micro Nikkor 105mm AF f/2.8, Micro Nikkor 105mm G AF-S/ED-IF/VR f/2.8, Sigma Macro 105mm AF/EX/DG/OS/HSMS f/2.8). Floral details were captured with a Micro-Nikkor 60mm f.2.8 mounted on a Nikon PB6 bellow or on extension tubes, and/or with microscopes (Leica macroscope Z16APO and Leica dissecting stereo microscopes MZ9.5 and M60) fitted with Leica DFC420 Digital Color Microscope cameras. Comparative plates of flowers and/or relevant floral details were prepared with Adobe® Photoshop 5.0 and 6.0. Drawings of flowers and floral details were prepared using dissecting stereo microscopes Leica MZ9.5 and M60, fitted with Leica ApoPlanar lenses and with drawing tubes. One or more vouchers for each specimen were conserved in the liquid collection of JBL. Individual plants intended for the description of new species were also preserved as dried specimens to be deposited in the herbaria of the National Museum of Costa Rica (CR) and the University of Costa Rica (USJ). Measurements were mostly taken under a dissecting stereoscope, or with the aid of the electronic scale bars inserted in the high definition images of the floral details. The major herbaria of the country, as well as relevant herbaria that allow digital access to their collections, were reviewed to study specimens of the taxa intended for this study, and specimens annotated when required.

DNA extraction and sequencing.— The selection of the material was based on availability and inter-specific variability. Whenever possible species are represented by more than a single accession (Table 1). Vouchers of the specimens are kept in the liquid collections at JBL or L, unless specified otherwise. Fresh leaf and flower cuttings of about 1 cm² were obtained from the selected individuals of each species. Each individual sample was put into a polypropylene bag with silica gel to dry for about a week after which the silica was removed and new dry silica was added. Twenty-mg samples were pulverized in liquid nitrogen by shaking them in a Retsch MM 300 shaker for 5 min using three bullets. Extraction was performed following the DNEasy extraction

TABLE 1. List of accessions used in the phylogenetic analyses here presented. The vouchers, NCBI GenBank accession number and source are given.

Taxon	Voucher	ITS	Source
<i>Arpophyllum giganteum</i> Hartw. ex Lindl.	None	AF266742	Pridgeon et al 2001
<i>Barbosella cucullata</i> (Lindl.) Schltr.	Chase 1334	AF262815	Pridgeon et al 2001
<i>Barbosella handroi</i> Hoehne	Chase 1436	AF262813	Pridgeon et al 2001
<i>Barbosella miersii</i> (Lindl.) Schltr.	Chase 6809	AF262816	Pridgeon et al 2001
<i>Barbosella orbicularis</i> Luer	Chase 1444	AF262814	Pridgeon et al 2001
<i>Dresslerella elvallensis</i> Luer	Chase 1279	AF262901	Pridgeon et al 2001
<i>Dresslerella hirsutissima</i> (C.Schweinf.) Luer	Chase 1306	AF262902	Pridgeon et al 2001
<i>Dresslerella pertusa</i> (Dressler) Luer	Chase 1326	AF262903	Pridgeon et al 2001
<i>Echinosepala aspasicensis</i> (Rchb.f.) Pridgeon & M.W.Chase (1)	Bogarín 8211	KP012468	This Study
<i>Echinosepala aspasicensis</i> (Rchb.f.) Pridgeon & M.W.Chase (2)	Bogarín 1945	KP012469	This Study
<i>Echinosepala aspasicensis</i> (Rchb.f.) Pridgeon & M.W.Chase (3)	Chase 971	AF262905	Pridgeon et al 2001
<i>Echinosepala expolita</i>	Bogarín 1871	KP012479	This Study
<i>Echinosepala longipedunculata</i> (1)	Bogarín 5449a	KP012480	This Study
<i>Echinosepala longipedunculata</i> (2)	Bogarín 5449b	KP012481	This Study
<i>Echinosepala pan</i> (Luer) Pridgeon & M.W.Chase (2)	Bogarín 1913	KP012471	This Study
<i>Echinosepala sempergemmata</i> (Luer) Pridgeon & M.W.Chase (1)	Bogarín 7137	KP012472	This Study
<i>Echinosepala sempergemmata</i> (Luer) Pridgeon & M.W.Chase (2)	Bogarín 5775	KP012473	This Study
<i>Echinosepala sempergemmata</i> (Luer) Pridgeon & M.W.Chase (3)	Bogarín 10717	KP012474	This Study
<i>Echinosepala stonei</i> (Luer) Pridgeon & M.W.Chase (1)	Bogarín 5777a	KP012482	This Study
<i>Echinosepala stonei</i> (Luer) Pridgeon & M.W.Chase (2)	Bogarín 5777b	KP012483	This Study
<i>Echinosepala stonei</i> (Luer) Pridgeon & M.W.Chase (3)	Bogarín 7190	KP012484	This Study
<i>Echinosepala tomentosa</i> (Luer) Pridgeon & M.W.Chase	Bogarín 5622	KP012485	This Study
<i>Echinosepala uncinata</i> (Fawc.) Pridgeon & M.W.Chase (1)	Chase 1321	AF262904	Pridgeon et al 2001
<i>Echinosepala uncinata</i> (Fawc.) Pridgeon & M.W.Chase (2)	Bogarín 5871b	KP012487	This Study
<i>Echinosepala vittata</i> (Pupulin & M.A.Blanco) Luer	Blanco 1324	KP012487	This Study
<i>Myoxanthus exasperatus</i> (Lindl.) Luer	Chase 1323	AF262882	Pridgeon et al 2001
<i>Myoxanthus punctatus</i> (Barb.Rodr.) Luer	Chase 1324	AF262885	Pridgeon et al 2001
<i>Myoxanthus serripetalus</i> (C.Schweinf.) Luer	Chase 1329	AF262883	Pridgeon et al 2001
<i>Pleurothallopsis nemorosa</i> (Barb.Rodr.) Porto & Brade	Bock s.n.	AF291099	Pridgeon et al 2001
<i>Pleurothallopsis striata</i> (Luer & R.Escobar) Pridgeon &	Chase 1103	AF262910	Pridgeon et al 2001
<i>Restrepia antennifera</i> Kunth	Chase 982	AF262906	Pridgeon et al 2001
<i>Restrepia aristulifera</i> Garay & Dunst.	Chase 1109	AF262907	Pridgeon et al 2001
<i>Restrepia muscifera</i> (Lindl.) Rchb.f. ex. Lindl.	Chase 308	AF262908	Pridgeon et al 2001
<i>Restrepiella ophioccephala</i> (Lindl.) Garay & Dunst.	Chase 291	AF262909	Pridgeon et al 2001

protocol (QIAGEN). DNA concentration for each sample was adjusted to 10 µmol/l using a Nano Drop Spectrophotometer (ND 1000). The nuclear ribosomal internal transcribed spacer (ITS) region was amplified using the methods and primers 17SE (ACGAATTCATGGTCCGGTGAAGTGTTCG) and 26SE (TAGAATCCCCGGTTCGCTCGCCGTTAC) for sequencing and amplification, as described

by Sun *et al.* (1994). Amplification was done by preparing each sample with a PCR mix composed of DTB, dNTPs, both primers (four in the case of ITS), Dream Taq, water, and the extracted DNA. Samples were amplified in a MJ Research PTC-200 Pelthier Thermal Cycler, using a temperature profile of 94°C/5 min, followed by 34 cycles of 94°C/30 s, 55°C/30 s, and 72°C/2 min, and finally 72°C/10 min. To prepare

for sequencing, a DETT kit was used according to the manufacturer's specifications (GE Healthcare). Each sample had two mega-mixes, one for the F-primer and another for the R-primer (four for ITS), and were cycle-sequenced using a standard thermo-profile of 94°C/20 s, 50°C/15 s, and 60°C/1 min, repeated 25 times. Products were analyzed on an ABI 9600 DNA analyzer.

Building the data sets.— The STADEN (Staden *et al.* 2003) package was used for editing the sequences. Where more than one base pair was equally probable, the Unicode nomenclature (IUPAC) was used. In a few cases the two traces for one sample were too short and there was no overlap so Pregap was unable to build a contig. In order to keep the information, both sequences were merged by filling in missing positions with N's. Sequences were aligned manually in Mesquite v2.72 (Maddison & Maddison 2007). The ends of each data set were trimmed to eliminate possible erroneous data, and gaps at the ends of sequences were regarded as missing data (filled in with N's). Each indel and possible informative sites were re-checked by going back to the original traces. After the alignments had been edited, additional sequences were obtained from GenBank, the latter using nBLAST. *Arpophyllum giganteum* Hartw. ex Lindl. AF266742 (ITS) was used as outgroup in all cases, as it is suggested to be the furthest related of all included species (Pridgeon *et al.* 2001).

Phylogenetic analysis.— Bayesian analyses methods were preferred over Parsimony and Maximum Likelihood because they were found to explain the data better and have overall higher support and resolution. The Bayesian Evolutionary Analysis Sampling Trees (BEAST; Drummond & Rambaut 2007) was used to analyze the ITS matrix. Substitution and clock models were unlinked. GTR + Γ model, estimated frequencies, and 8 categories were used. The Lognormal relaxed clock model was used. The tree prior was speciation - Yule birth, while the number of generations was set to 30,000,000. The first 20% of the resulting trees were used as burnin. Trees were visualized in FigTree v.1.3.1 (Rambaut 2009). Posterior probability (PP) values were added to the branches of the trees using the labeling option. Branches were reordered for better visualization.

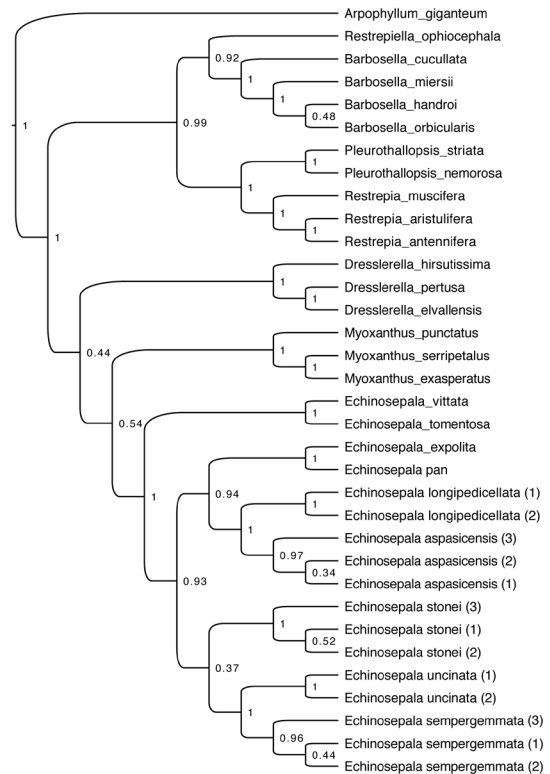


FIGURE 7. Phylogenetic relationship amongst the species of *Echinosepala*. The trees were produced with an analysis of ITS datasets, using BEAST v1.8.0. Parameters were set to preset, except for substitution model GTR with 8 categories, clock model uncorrelated lognormal (ITS), tree prior Yule process, and number of generations 30,000,000. The resulting trees were combined using TreeAnnotator v1.6.0., with the first 20% of the trees were used as burnin. Node values are posterior probabilities. Tree edited using FigTree v.1.3.1.

Results. Species of *Echinosepala* form a highly supported clade, consecutively sister to *Myoxanthus*, *Dresslerella*, and a clade grouping species of *Pleurothallopsis* and *Restrepia*, and species of *Restrepiella* and *Barbosella* on two branches (Fig. 7). Within *Echinosepala*, *E. aspasicensis* and *E. longipedunculata* are highly supported as sister species, and in turn they are well supported as sister to *E. pan* and *E. expolita*. The latter two species cannot be differentiated by their ITS sequences.

Echinosepala sempergemmata and *E. uncinata* form a well-supported group, which in turn is sister (with low support) to *E. stonei*. In our analysis, the

phylogenetic position of *E. stonei* is not resolved, but the species is surely not closely related to the group of *E. expolita/E.pan*. In order to better resolve the relationships between the derived species with a smooth lip callus and no pseudo-glenion, the analysis would surely benefit by including a sample of *E. lappiformis*, which is very close to *E. stonei* in both vegetative and flower morphology.

Echinosepala tomentosa and *E. vittata*, both characterized by miniature plant habits and flowers, are sister to all the other species in a highly supported group.

NEW TAXA

Echinosepala expolita Pupulin & Belfort, *sp. nov.*

TYPE: Costa Rica. Alajuela: San Ramón, Piedades, Piedades Norte, road to Bajo La Paz, ca. km 3, along the Río San Pedro, 10°08'58.7"N 84°34'03.3"W, 1300 m, premontane wet forest, secondary and remnants of primary vegetation, 19 February 2008 *F. Pupulin 7030*, *R.L. Dressler & A. P. Karremans* (holotype, JBL!; isotypes, JBL!). Fig. 1C, 2, 3B, 4D, 8–13.

Species Echinosepala pan (Luer) Luer *similis*, *caulum vaginis glabris paulo inflatis, foliis ligulato-lanceolatis multo latioribus, ovario lanato-hirsuto, floribus multo majoribus indumento abaxialis sepalorum tomentoso munitis praecipue differt.*

Epiphytic, caespitose, erect *herb* up to 30 cm tall. *Roots* coarse, flexuous, 1.5–2.0 mm in diameter. *Ramicauls* stout, erect, terete, thicker in the distal portion, 6.0–12.5 cm long, homoblastic, composed by 5 nodes different in length, completely enclosed by 3–5 papyraceous, tubular, ancipitous, apically loose, obliquely truncate, whitish, glabrous, inflated sheaths, increasing in size toward the upper one, 1.4–7.7 × 0.2–0.8 cm, the oldest ones breaking longitudinally and eventually disintegrating with age. *Leaf* erect, coriaceous, linear-elliptic, subacute, minutely emarginate at the apex, 1.8–2.4 × 16.2–27.0 cm, with the midvein strongly protruding abaxially. *Inflorescence* a fascicle of single, successive flowers produced facultatively at the apex and the base of the ramicaul at once; the peduncle terete-subclavate, erect, glabrous, 1.8–2.5 mm long. *Floral bract* papyraceous, loose, obliquely truncate-subobtuse, 1.8–2.4 cm long.

Pedicel terete-subclavate, 1.8–2.5 cm long; *ovary* lanate-hirsute, completely covered by the bract, linear-subclavate, 3 mm long. *Flowers* bilabiate, with temporal activity, fully opening during the morning and closing in late afternoon; the sepals light greenish yellow, with the adaxial basal half mottled purple-red and the distal half with pointed verrucae becoming denser toward the apex, abaxially flushed with purple and densely warty-pubescent; the petals yellow, blotched with purple-red, sparsely verrucose; the lip dark purple, apically covered with a translucent wax in fresh flowers. *Dorsal sepal* elliptic-oblong with apex subacute, 1.9–2.0 × 0.4–0.9 cm, 7- to 9-veined, a semitransparent base forming a window under the dark purple-red blotches, tomentose abaxially, adaxially provided with spiny warts in the apical half. *Lateral sepals* connate into an elliptic, abaxially tomentose synsepal, 1.6–1.7 × 0.4–1.0 cm, each sepal 5-veined, the apices free about 6 mm, subacute-rounded, with pointed-spiny warts in the distal half. *Petals* fleshy, rhombic, obliquely asymmetric, acute, 6–7 × 1–3 mm, 3-veined, apically provided with spiny warts. *Lip* 3-lobed, narrowly oblong-obovate from a small, rounded, thin, dark purple claw, 7.5 × 2.2 mm (5.0 mm across the lateral lobes expanded), the base subtruncate; lateral lobes basal, erect, narrowly linear-subuncinate, antrorse; the apical lobe elliptic-truncate, with irregular margins, provided with a pair of intramarginal, thin keels running from the disc, inside the lateral lobes, to the middle of the midlobe blade, thickened at the apex into a cushion-like, elliptic, low pad; the disc with an erect, narrow callus arising above the base the lip, covered on the top by a horseshoe-shaped, channeled pseudo-glenion, from which a gelatinous exudate is produced, extending in front into a low groove flushing into the apical pad. *Column* straight to subarcuate, semiterete, 4.4–5.5 mm long, provided with long, narrow, subrectangular wings above the middle, the foot ca. 2.3 mm long. *Anther cap* globose, cucullate, with conspicuous, stiff hairs on the upper margin, 2-celled. *Pollinia* 2, obovoid, flattened, on a short bilobed caudicle. *Capsule* not seen.

PARATYPE: Costa Rica. Alajuela: San Ramón, Ángeles, Reserva Biológica Alberto M. Brenes, 10°13'06" N 84°36'11" W, 850 m, bosque muy húmedo tropical transición a premontano, sobre el Sendero La Catarata,

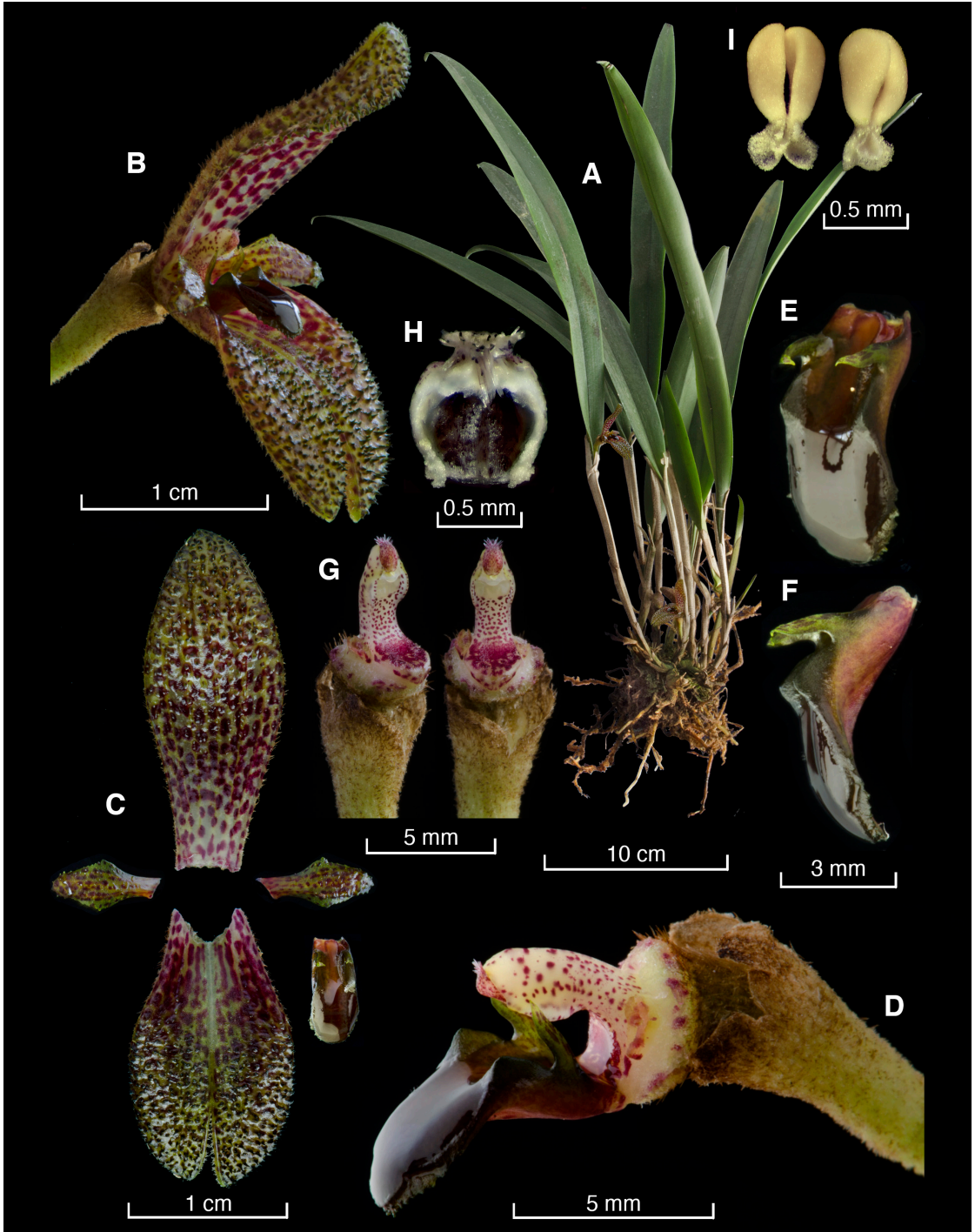


FIGURE 8. Lankester Composite Digital Plate of *Echinosepala exposita*. A. habit. B. flower. C. dissected perianth. D. ovary, column and lip in lateral view. E–F, labellum, three-quarters and lateral views. G. column, three-quarters and ventral views. H, anther cap. I, pollinarium, dorsal and ventral views. Based on the holotype. Photographs and digital composite by F. Pupulin.

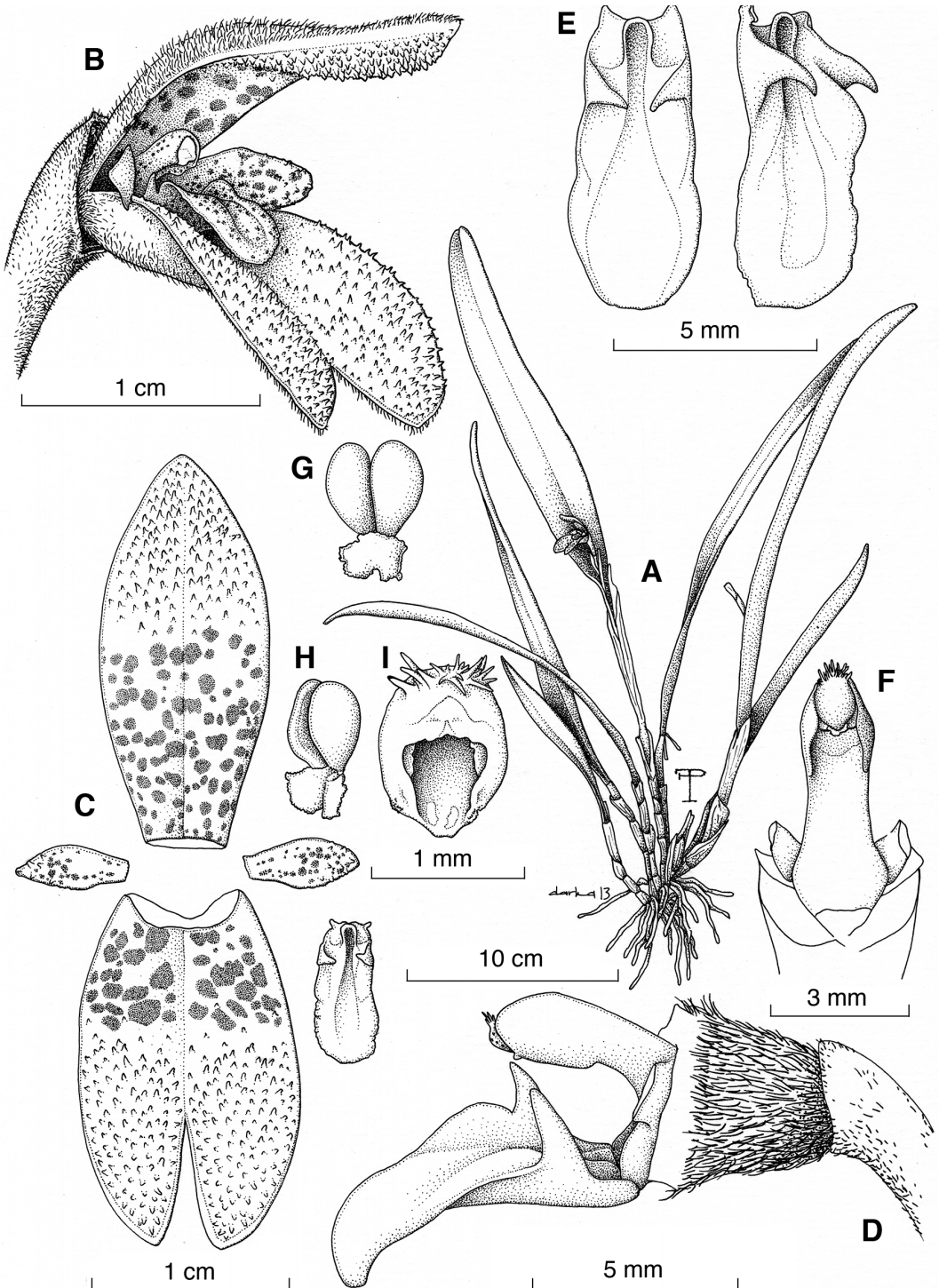


FIGURE 9. *Echinosepala expolita* Pupulin & Belfort. A. habit. B. flower. C. dissected perianth. D. ovary, column and lip in lateral view. E. labellum in ventral and three-quarters views. F. column, ventral view. G–H. pollinarium in dorsal and three-quarters views. I. anther cap. Based on the holotype. Drawing by F. Pupulin and D. Solano Ulate.



FIGURE 10. Flower of *E. expolita*, from the plant that served as the holotype. Photograph by F. Pupulin.



FIGURE 11. Flower of *E. expolita* (Bogarín 1871). Photograph by F. Pupulin.

epífitas en borde de bosque secundario a orillas del Río San Lorencito, 24 setiembre 2005, *D. Bogarín 1871* (USJ; isoparatype, JBL!) (Fig. 11).

OTHER MATERIAL EXAMINED: Costa Rica. [Alajuela:] Piedades S. de San Ramón, Cerros de Laguna, 1190 m, 7 Nov.1922, *A. M. Brenes 26219 (425 Herb. Brenes)* (CR!, sterile); Alajuela: San Ramón, Cordillera de Tilarán, cerca a Balsa, ca. 12 km NE de San Ramón, 10°10'30" N 80°30'30" W, 1100 m, 18 June 1995. Epífita a 2 m. Flores gris con manchas moradas y rayas blanco-hueso, suculentas. *B. Hammel 19870* (CR!, sterile).

ETYMOLOGY: From the Latin *expolitus*, “shining, glossy,” in allusion to the glossy apex of the lip in the fresh flowers with exudate.

DISTRIBUTION: Only known from the Caribbean watershed of the Cordillera de Tilarán in central Costa Rica (Fig. 14).

HABITAT AND ECOLOGY: Epiphytic in tropical, transition to premontane, and premontane wet forests along the Caribbean watershed of the Cordillera de Tilarán, at 800-1300 meters of elevation. Flowering has been recorded from March to June, and in November, but it is probable that the plants may flower at any time of the year. Each individual flower shows temporal activity, fully opening during the morning and closing in late afternoon. Flowers are usually almost to completely closed at night. During anthesis they emit a subtle but pungent smell, reminiscent of urea. The breakdown of urea to ammonia and carbon dioxide is known to lure insects, particularly fruit flies.

In plant architecture, *E. expolita* is similar to *E. aspasicensis*, with narrowly lanceolate leaves longer than the subtending ramicauls. The inflorescences, however, may be produced both at the apex of stem and from one of the lower nodes of the ramicaul, and may be exerted simultaneously (Fig. 2). The basal

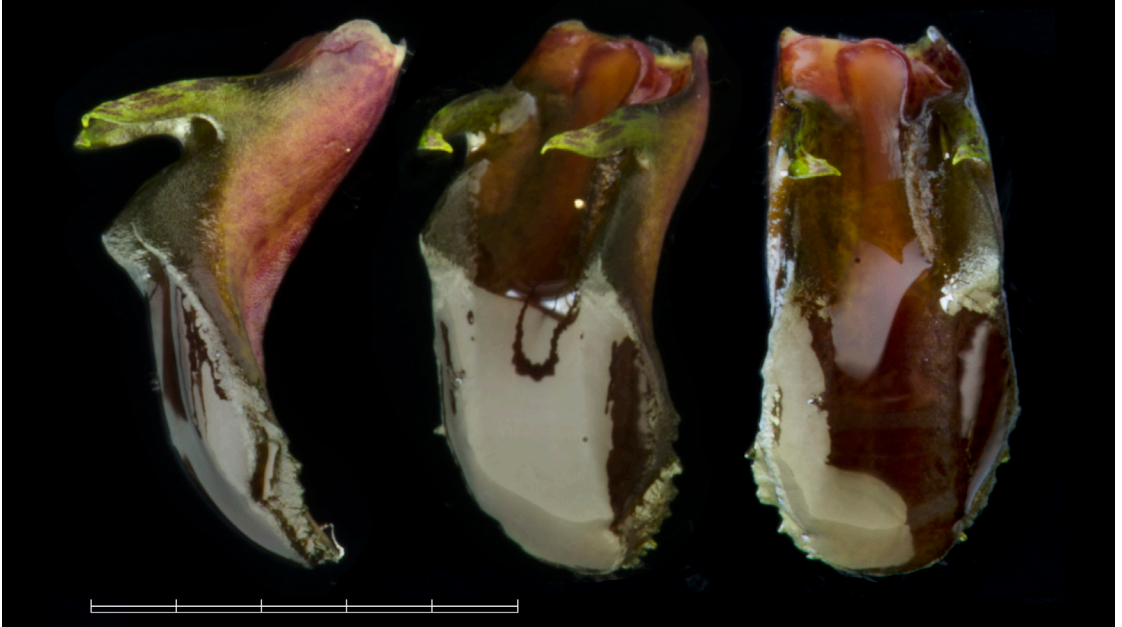


FIGURE 12. Three views of the labellum of *E. expolita* (Pupulin 7030) from a recently open flower. Note the shiny surface of the lip blade. Photographs by F. Pupulin.



FIGURE 13. Comparison between the flowers of *E. expolita* (A) and *E. pan* (B). A, Pupulin 7030. B, Bogarín 10270. Vouchers at JBL. Photographs by F. Pupulin.

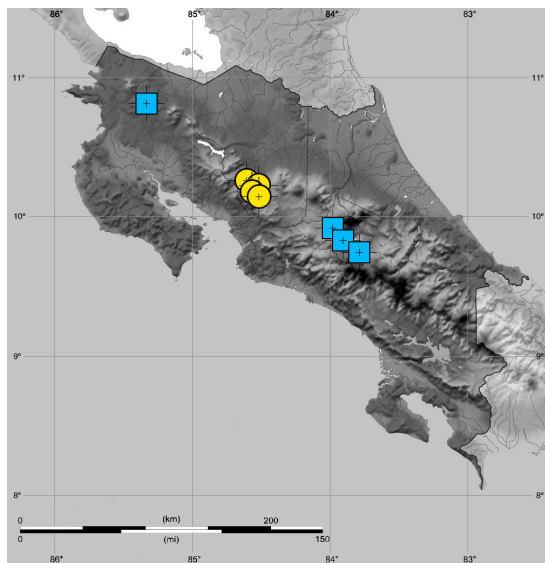


FIGURE 14. Distribution map of *Echinosepala expolita* (yellow circles) and *E. longipedunculata* (blue squares) in Costa Rica.

inflorescences are apparently not an ancestral condition in the group (Fig. 7). They may have originated on two independent occasions (in the clades of *E. expolita*+*E. pan* and in the clade of *E. stonei*). It is also possible that it evolved once, and one or two reversals to the ancestral state (apical inflorescence) occurred. Either scenario indicates that the species with basal inflorescences do not form a monophyletic group in *Echinosepala*.

Florally, *E. expolita* is close to *E. pan*, to which it appears related in the phylogenetic reconstruction presented (Fig. 7), and from which it can be distinguished by the glabrous, inflated sheaths that cover the stem (vs. hirsute, tight), the distinctly broader linear-elliptic leaves (vs. narrowly lanceolate), the lanate-hirsute ovary (vs. echinate), the much larger flower (sepals >15 mm vs. <10 mm long), and the tomentose abaxial indumentum of the sepals (vs. hirsute-echinate). At anthesis and in fresh flowers, the adaxial surface of the lip is covered with a translucent, waxy substance, which makes the lip shiny the basis for the specific epithet) and seemingly wet. This exudate, which appears to spread from the basal pseudo-glenion over the lip blade, is not sticky to touch, and it becomes progressively matte until the lip appears dry during the last days of anthesis.

The pseudo-glenion is apparently an ancestral character in *Echinosepala*, as it has been recorded in both the *E. vittata*+*E. tomentosa* clade, and in the

clade including *E. aspicensis*+*E. longipedunculata* and *E. expolita*+*E. pan*, and it was lost in the clade(s) of *E. lappiformis*+*E. stonei* and *E. sempergemmata*+*E. uncinata*.

It is noteworthy that despite the overwhelming differences in both vegetative architecture and flower morphology between the two species, *E. expolita* and *E. pan* have not acquired enough visible changes in the analyzed ITS region to be differentiated by genetic analysis. A very low nrITS sequence divergence in morphologically well distinguished but closely related species was also reported by Karremans *et al.* (2015) in genus *Specklinia* Lindl.

***Echinosepala longipedunculata* Pupulin & Karremans, sp. nov.**

Echinosepala aspicensis (Rchb.f.) Pridgeon & M.W.Chase *plerumque similis*, *pedunculo pro genere longissimo, floribus viridiflavis* (vs. *brunneo-purpureos*), *sepalorum indumento adaxiali verrucosotuberculato* (vs. *glabrum*), *labello aurantiace truncate brevior* (vs. *unum dimidiumque longiore, obtusum vel acutum, purpureum*) *recedit*.

TYPE: COSTA RICA. Cartago: La Unión, San Rafael, Cerros de La Carpintera, Campamento Escuela Iztarú, 9°53'08.2"N 83°58'15.6"W, 1778 m, bosque húmedo premontano, epífitas en potreros arbolados, 30 octubre 2008, floreció en cultivo en el Jardín Botánico Lankester, 19 de junio 2012, *D. Bogarín 5449, R.L. Dressler, R. Gómez, F. Pupulin, & R. Trejos* (holotype, JBL!). Fig. 4F–4H, 15, 16A, 17A.

Epiphytic, caespitose, erect *herb* up to 30 cm tall. *Roots* coarse, flexuous, 1.5–2.0 mm in diameter. *Ramicauls* stout, erect, terete, slightly complanate, thicker in the distal portion, 5.5–10 cm long, homoblastic, composed of 2 nodes differing in length, the lower one much shorter (to 1.5–2.2 cm long), completely enclosed by 3 papyraceous, tubular, ancipitous, apically loose, obliquely truncate, whitish sheaths, increasing in size toward the upper one, 2.0–6.5 × 1.3–1.8 cm, the oldest ones breaking longitudinally into long fibres and eventually disintegrating with age. *Leaf* erect, thickly coriaceous, linear-elliptic, subacute, minutely emarginate at apex, 15–18 × 2.3–3.0 cm, cuneate below into a conduplicate, sessile base, with the midvein strongly

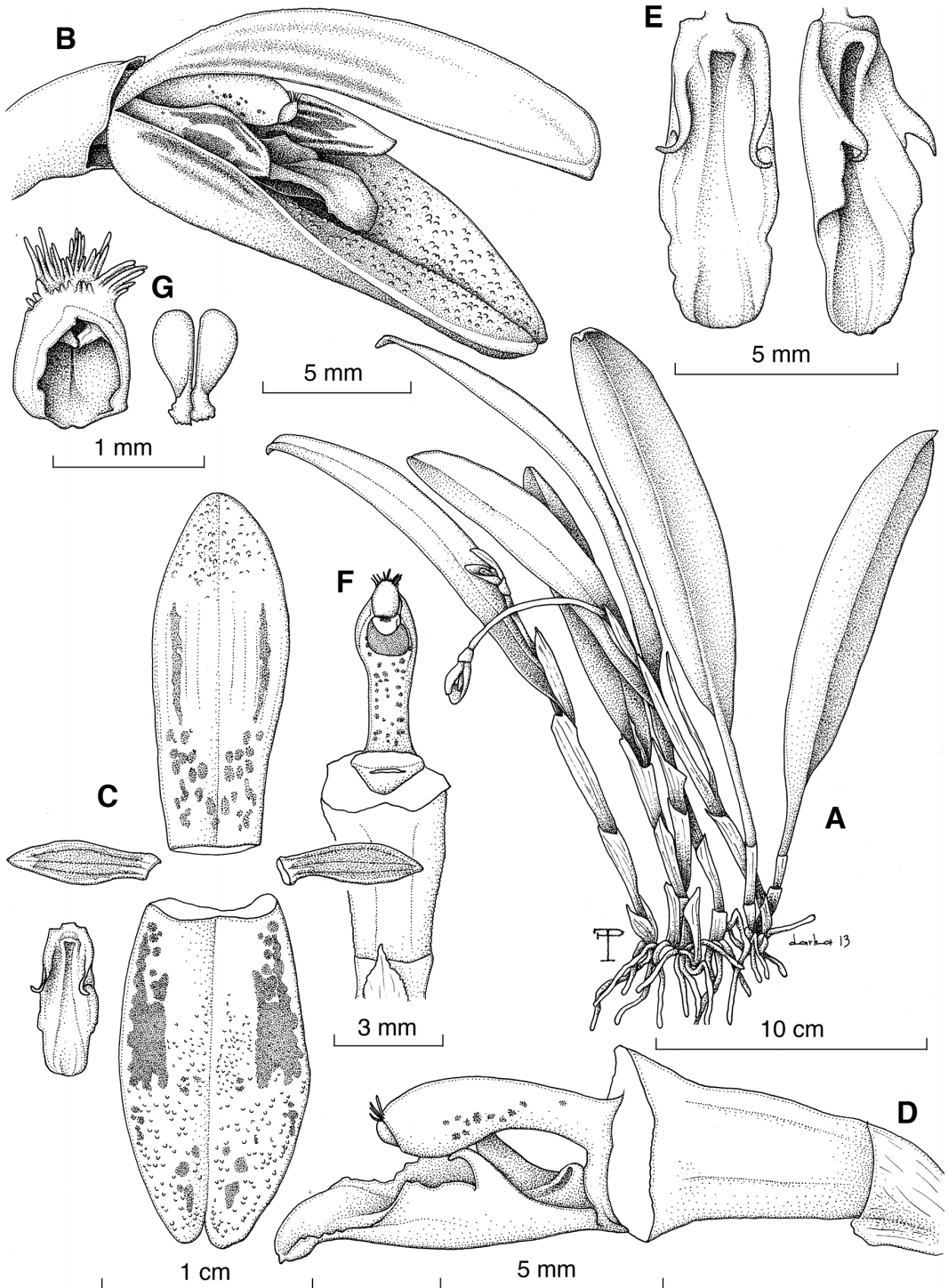


FIGURE 15. *Echinosepala longipedunculata* Pupulin & Karremans. A. habit. B. flower. C. dissected perianth. D. ovary, column and lip in lateral view. E. labellum in ventral and three quarters views. F. column, ventral view. G. anther cap and pollinarium. Based on the holotype. Drawing by F. Pupulin and D. Solano Ulate.

protruding abaxially and several lateral veins faintly visible on the adaxial blade. *Inflorescence* a fascicle of single, successive flowers at the apex of the ramicaul; the peduncle terete, suberect to gently arching, sparsely and minutely pubescent, 3–8 cm long, subtended by a papyraceous, whitish, ancipitous, obliquely truncate spathe 2.7–4.2 cm long. *Floral bract* papyraceous, loose, conduplicate, obliquely truncate-subobtuse, 7–10 mm long. *Pedicel* terete-subclavate, 4–5 mm long; *ovary* completely covered by the bract, linear-subclavate, 5 mm long, minutely scaberulous. *Flowers* bilabiate, with temporal activity, opening early in the morning and closing in the afternoon for three to four days; the sepals yellow, mottled dark purple on the inner side, flushed purple and densely scabrous to shortly pubescent externally, adaxially verrucose, the verrucae scattered at the base and becoming denser toward the apex, the petals yellow, heavily blotched with purple-red, the lip orange-yellow. *Dorsal sepal* oblong-obovate, obtuse, 19–21 × 6–7 mm, 7- to 9-veined. *Lateral sepals* connate into an elliptic synsepal, verruculose in the distal half, 17–19 × 9–10 mm, each sepal 5-veined, the free apices subacute-rounded. *Petals* fleshy, narrowly lanceolate, obliquely asymmetric, acute, 6–7 × 1.5–1.8 mm, 3-veined, the central vein thickened towards the base into a rounded keel. *Lip* 3-lobed, narrowly elliptic from a small, rectangular, thin, hyaline claw, 7 × 2 mm (4.5 mm across the lateral lobes expanded), the base subtruncate; lateral lobes erect, narrowly uncinuate, antrorse; the apical lobe rounded to subtruncate, thickened at the apex into a cushion-like, elliptic, low pad, provided with a pair of intramarginal, thin keels running inside the lateral lobes toward the disc; the disc with an erect, narrow callus arising above the base of the lip, the highest part covered by a horseshoe-shaped, channeled pseudo-glenion extending in front into a narrow groove flushing and expanding into the apical pad. *Column* straight to subarcuate, semiterete, 5.0–5.5 mm long, provided with narrow, broadly rounded wings above the middle, the foot ca. 1.5 mm long. *Anther cap* globose, cucullate, with conspicuous, stiff hairs on the upper margin, 2-celled. *Pollinia* 2, obovoid, flattened, on a short, bilobed caudicle.

PARATYPES: COSTA RICA. Cartago: Cartago, San Francisco, Muñeco, Finca Loma Verde y Jilguero,

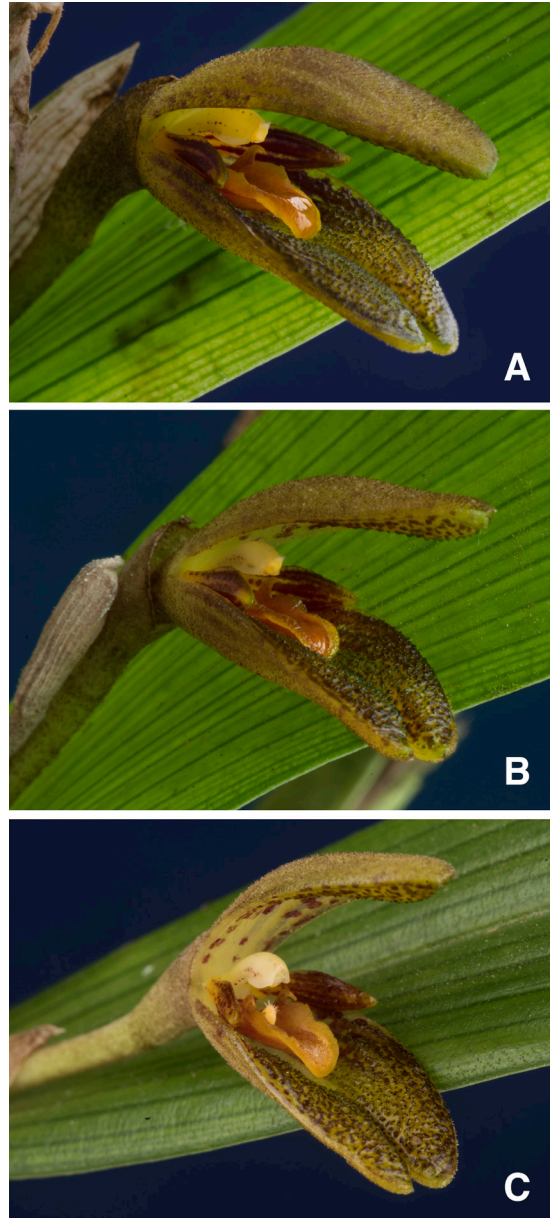


FIGURE 16. Flowers of *Echinosepala longipedunculata*. A, Karremans 4944. B, Bogarín 5449. C, Chinchilla 2216. Vouchers at JBL. Photographs by F. Pupulin.

camino a Alto Belén, entre Río Sombrero y Quebrada Patarrá, 9°46'50.3"N 83°54'21.1"W, 1430–1620 m, bosque pluvial premontano, epifitas en bosque secundario y árboles en zonas abiertas, 23 mayo 2007, floreció en cultivo en el Jardín Botánico Lankester, 30 marzo 2013, D. Bogarín 3863, M. Bonilla, R. Gómez, Rafael Trejos & J.D. Zúñiga (JBL-spirit!); Cartago:



FIGURE 17. Comparison between the flowers of *E. longipedunculata* (A) and *E. aspicensis* (B). A, Bogarín 5449, Karremans 4407. Vouchers at JBL. Photographs by F. Pupulin.

Paraíso, Orosi, Tapantí, Parque Nacional Tapantí, unos 9 km después de la entrada principal del parque, 500 m después del puente superior sobre el río Grande de Orosi sobre la calle que va a la represa, 9°41'56.24"N 83°46'53.12" W, 1570 m, epífitas en árboles al lado de la calle principal, bosque pluvial premontano, 10 de febrero 2012, floreció en cultivo en el Jardín Botánico Lankester, 9 marzo 2013, *A.P. Karremans 4994* (JBL-spirit!) (Fig. 16B). Guanacaste: Liberia, Liberia. Parque Nacional Rincón de la Vieja, sendero que recorre el sector Volcán Santa María, 10°47'26.00»N, 85°19'16.00"O, 1304 m, bosque húmedo montano bajo, epífita, 24 Jan. 2015, flowered in cultivation at Lankester Botanical Garden, 29 Nov. 2016, *I. Chinchilla 2216*, *R. Espinoza*, *C. Moraga*, *A. Guadamuz & A. Rojas Alvarado* (JBL-spirit!) (Fig. 16C).

ETYMOLOGY: From the Latin *longipedunculatus*, "with a long peduncle," in reference to the unusually elongate peduncle of the flower, which can reach 8 cm in length.

DISTRIBUTION: Known only from Costa Rica, where

it has been recorded from the Caribbean watershed of the Talamanca, Central Volcanic, and Guanacaste mountain chains (Fig. 14).

HABITAT AND ECOLOGY: Epiphytic in partial shade on large branches and trunks of trees, in primary and mature secondary vegetation, with records in lower montane wet, premontane rain, and premontane wet forests at elevations of between 1300–1800 meters. Flowering has been recorded under cultivation in March to June, September, and November.

Echinosepala longipedunculata is most similar to *E. aspicensis*, with which it shares the large habit of the plant and the general morphology of the flower. Nevertheless, *E. longipedunculata* can be recognized by the unusually long peduncle that can reach 8 cm in length (vs. 2.0–3.5 cm in *E. aspicensis*), the greenish yellow flowers (vs. purple-brown), the verrucose-tuberculate adaxial surface of the sepals (vs. glabrous), and the shorter, truncate, orange lip (vs. one and half times longer, obtuse to acute, purple).

Whilst the phylogenetic reconstruction based on the ITS region clearly distinguishes between *E. longipedunculata* and *E. aspasicensis*, the samples of the latter taxon used in our analysis seem to suggest that the concept of *E. aspasicensis* encompasses two or more cryptic species.

KEY TO THE SPECIES OF *ECHINOSEPALA* IN COSTA RICA

1. Base of the lip with the callus with a horseshoe-shaped pseudo-glenion 2
2. Inflorescences produced from the lower nodes of the ramicaul 3
 3. Sheaths of the ramicaul hirsute; sepals echinate abaxially *E. pan*
 - 3a. Sheaths of the ramicaul glabrous; sepals shortly tomentose abaxially *E. expolita*
- 2a. Inflorescences produced from the apex of the ramicaul 4
 4. Leaves of fertile stems < 10 long; flowers small, synsepal < 1 long, lip < 4 mm long 5
 5. The dorsal sepal yellow, the synsepal deep purple, abaxially densely tomentose *E. tomentosa*
 - 5a. Sepals whitish, striped with red, abaxially with stiff hairs along the veins *E. vittata*
 - 4a. Leaves of fertile stems > 15 long; flowers large, synsepal > 15 mm long, lip > 7 mm long 6
 6. Ovary and abaxial surface of the sepals tomentose; adaxial surface spiny *E. expolita*
 - 6a. Ovary and abaxial surface of the flowers warty; adaxial surface verruculose 7
 7. Peduncle < 3 cm long; flowers purple-maroon; lip elliptic, acute, purple-red *E. aspasicensis* s.l.
 - 7a. Peduncle > 6 cm long; flower yellow-orange; lip sub rectangular, truncate, yellow *E. longipedunculata*
- 1a. Base of the lip with the callus smooth to rugulose, without a pseudoglenion 8
 8. Inflorescences produced from the lower nodes of the stem 9
 9. Dorsal sepal narrowly triangular-lanceolate *E. lappiformis*
 - 9a. Dorsal sepal broadly oblong *E. stonei*
 - 8a. Inflorescence produced from the apex of the stem 10
 10. Leaves ligulate; flowers autogamous, mostly cleistogamous, adaxially warty *E. sempergemmata*
 - 10a. Leaves lanceolate; flowers not autogamous, adaxially glabrous *E. uncinata*

We take the opportunity of this paper to transfer to *Echinosepala* a species originally described by Luer (*Acianthera*, but clearly belonging in *Echinosepala* based on both its vegetative habit and flower morphology. Zelenko and Bermúdez (2009) recorded *E. aspasicensis* from Peru, without citing any voucher to support their claim. The present transfer, based on a specimen from the Department of Junín in the Amazonian region of Central Peru, fills the gap in the distribution of the genus *Echinosepala* noted by Pridgeon (2005a).

Echinosepala biseta* (Luer) Pupulin, *comb. nov.

Basionym: *Acianthera biseta* Luer, Harvard Papers in Botany 17: 333, f. 1. 2012.

TYPE: Peru. Junín, Chanchamayo, near Mina San Vicente, 2000-2800 m, June 2012, collected by Benito Monterrey, *E. Morón de Abad 004* (Holotype: USM).

ACKNOWLEDGMENTS. This manuscript was prepared as part of a dedicatory issue commemorating the ninety-fifth birthday of Carlyle A. Luer, whose extensive work on the Pleurothallidinae is the basis for most current studies in the subtribe. The scientific services of the Costa Rican Ministry of Environment and Energy (MINAE) and National System of Conservation Areas (SINAC) kindly issued the collecting permits under which wild specimens treated in this paper were collected. We are indebted to Diego Bogarín for the profound discussion on the taxonomy and systematic of the Pleurothallidinae. We thank Darha Solano Ulate for her help inking the illustrations of the new species. This research was enabled by the Project 814-A0-052, “Flora Costaricensis, Orchidaceae, subtribe Pleurothallidinae”, supported by the Vice-Presidency of Research, Universidad de Costa Rica.

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