Understanding trophic relationships among Caribbean sea urchins

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Abstract: The species Echinometra lucunter, Echinometra viridis, Lytechinus variegatus, Tripneustes ventricosus, and Diadema antillarum are the most common sea urchins of littoral habitats in the Caribbean. T. ventricosus and L. variegatus are associated with seagrass beds, while the other three species usually inhabit hardground substrates. Food preferences of these species are well documented and they are commonly accepted as being primarily herbivorous-omnivorous; nevertheless, few of them have previously been characterized isotopically. We used this approach for assessing the isotopic characterization of five echinoids. We established the trophic position of two groups of co-occurring species and quantified the contribution of food resources in the diet of Echinometra lucunter, considered the most common sea urchin in the Caribbean region. The species T. ventricosus and D. antillarum showed the highest values of δ15N. Sea urchins exhibited similar values of δ13C varying from -11.6 ± 0.63 to -10.4 ± 0.99%. The echinoid E. lucunter displayed the lowest values of carbon, from -15.40 ± 0.76%. Significant differences among species were found for δ15N and δ13C. Seaweed communities exhibited no differences among sites for overall δ15N (F= 1.300, df= 3, p= 0.301), but we found spatial differences for δ13C (F= 7.410, df= 3, p= 0.001). The ellipse-based metrics of niche width analysis found that the hardground biotope species (D. antillarum, E. lucunter, and E. viridis) did not overlap each other. Similar results were obtained for the co-occurring species of the seagrass biotope; however, the distance between these species was closer than that of the hardground biotope species. The Bayesian mixing models run for E. lucunter at all four localities found differences in food resources contribution. The algae Durvillaea menestralis, C. crassa and B. triquetra dominated in CGD; whereas C. nitens, Gracilaria spp., and D. caribaea represented the main contributor algae to the diet of E. lucunter at LQY. In Culebra Island, no dominance of any particular algae was detected in TMD, where six of the eight species exhibited a similar contribution. Similarities in δ15N between D. antillarum and T. ventricosus may hint towards a similar trophic level for these species, although T. ventricosus is widely accepted as an omnivore, while D. antillarum is considered a generalist herbivore. The lack of overlap among species in the two biotopes seems to indicate a resource partitioning strategy to avoid niche competition among co-occurring species. Rev. Biol. Trop. 64 (2): 837-848. Epub 2016 June 01.

Key words: stable isotopes, trophic ecology, Bayesian mixing models, sea urchins, Puerto Rico.
especially in marine ecosystems, which host a variety of species that occupy apparently similar trophic positions (McCutchan, Lewis, Kendall, & McGrath, 2003; Phillips, 2012). Co-occurring species generally coexist due to differences in feeding behavior and internal assimilation mechanisms (Wolf, Scott, & Martínez del Río, 2009). The presence of two or more species in the same area may be possible through the existence of resource partitioning, which serves to limit niche competition among sea urchins (Vanderklift, Kendrick, & Smith, 2006; Cabanillas-Terán, Loo-andrade, Rodríguez-Barreras, & Cortés, 2016).

Biological processes responsible for carbon and nitrogen movement in and out the ocean are critical in determining the natural abundance of these two elements in marine ecosystems (Nixon et al., 1996). However, there is a high degree of spatial variability in coastal marine systems, mainly in nearshore and continental shelf waters due to biotic and abiotic factors (Van Dover, Grassle, Fry, Garratt, & Starczak, 1992). Analysis of trophic relationships using stable isotopes is usually more complex, as primary producers and consumers are part of a complex trophic system in which changes in primary producers affect the isotopic signatures of herbivores (Fry & Sherr, 1984; Peterson, Howarth, & Garratt, 1985). For this reason, stable isotope studies have become a fundamental tool for assessing trophic niche interactions (DeNiro & Epstein, 1981; Caut, Angulo, & Courchamp, 2008; Parnell, Inger, Bearhop, & Jackson 2010; Boecklen, Yarnes, Cook, & James, 2011, Rodríguez-Barreras, Cuevas, Cabanillas-Terán, & Sabat, 2015).

Stable isotope analyses on dietary reconstructions rely on tissue signatures that provide valuable information on trophic level and potential food source choice in the diet of a given organism (Hobson & Sease, 1998; McCutchan et al., 2003; Vanderklift & Ponsard, 2003; Caraveo-Patiño & Soto, 2005). Thus, carbon and nitrogen stable isotopes have been used to elucidate upon food habits (Peterson & Fry, 1987), including the source and transformation of ingested organic matter (Peterson, Howarth, & Garratt, 1985), as well as on nutrient migration within food webs, trophic positioning of organisms, and their contribution to each trophic level (Vander-Zanden & Rasmussen, 1996). Additionally, these analyses have provided evidence for high densities of individuals of different species occupying the same trophic position in the same ecosystem (Vanderklift et al., 2006). Mathematical algorithms have developed to estimate the contribution of each food resource to the consumer, providing valuable information towards their hierarchical contribution (Solomon et al., 2011). Known as mixing models, these routines have been used extensively as powerful tools in trophic studies (Semmens, Moore, & Ward, 2009; Phillips, 2012). Mixing models have clarified important aspects of diet plasticity, differential assimilation of resources, association with biosynthetic pathways, and ecological consequences among others (Villinski, Hayes, Villinski, Brassell, & Raff, 2004; Tomas, Alvarez-Cascos, Turon, & Romero, 2006; Wing, McLeod, Clark, & Frew, 2008). Furthermore, the most recent Bayesian ellipses techniques have allowed stronger comparisons of isotopic niches across communities (Jackson, Inger, Parnell, & Bearhop, 2011).

Sea urchin digestive processes may be considered among the best studied among marine invertebrates (Lewis, 1964; Trenzado et al., 2012; Lawrence, Lawrence, & Watts 2013). For instance, Klinger, Hsieh, Pangallo, Chen, & Lawrence (1986) and Hawkins (1981) studied assimilative processes in *L. variegatus* and *D. antillarum*, respectively. Additionally, there are several studies of echinoids using a stable isotope approach (Minagawa & Wada, 1984; Rodriguez, 2003; Tomas et al., 2006; Vanderklif et al., 2006; Wing et al., 2008; Prado, Alcoverro & Romero, 2010; Wangensteen et al., 2011). Specifically, Cabanillas-Terán (2009) quantified its food resources contribution in the diet of *Diadema af. africanum*, formerly *Diadema antillarum*; whereas Prado, Carmichael, Watts, Cebria & Heck, (2012) experimentally determined the trophic enrichment factor for the green echinoid *Lytechinus variegatus*. Despite this, there is no other available information for
sea urchins in the Caribbean related to their trophic interactions or diet assimilation of food resources using stable isotopes. We used this approach for assessing the isotopic characterization of five echinoids. We established the trophic position of two groups of co-occurring species and quantified the contribution of food resources in the diet of *Echinometra lucunter*, considered one of the most common sea urchin in the Caribbean region.

**MATERIAL AND METHODS**

**Study site and data collection:** Surveys were conducted in October of 2013 at four reef crests of the Puerto Rico Archipelago. Two sites were located on the Northeastern coast of Puerto Rico, with the other two located off the island of Culebra (Fig. 1). Sites were Cerro Gordo (CGD: 18°16'51.40" N - 65°17'12.21" W), Luquillo (LQY: 18°23'18.46" N - 65°43'5.52" W), Tamarindo Bay (TMD: 18°18'55.29" N - 65°19'5.82" W), and Melones (MLN: 18°18'15.70" N - 65°18'42.27" W). All sites are shallow-water fringing reefs between 1-2m in depth. Culebra Island has no rivers, precipitation average is low, and volcanic and intrusive rocks, with some limestone deposits (U.S.G.S., 1996), characterize the substrate. The sites CGD and LQY are composed of carbonate rocks (Rodríguez-Barreras, Pérez, Williams, Mercado-Molina, & Sabat, 2014).

We collected five adults of the species *Diadema antillarum*, *Echinometra viridis*, *Lytechinus variegatus*, and *Tripneustes ventricosus* from LQY. Additionally, five individuals of *Echinometra lucunter* were collected from all four localities (Fig. 1). The genus *Echinometra* and *D. antillarum* were collected in hardground biotopes (fringing reef), whereas *L. variegatus* and *T. ventricosus* were gathered in a backreef lagoon biotope covered by seagrass meadows. Sample sizes were based on previous studies (Rodriguez, 2003; Tomas et al., 2006; Wing et al., 2008; Cabanillas-Terán, 2009). Thus, we collected samples of all seaweed species within a 5m radius from collecting sites associated only with *E. lucunter*. All organisms, including algae and sea urchins, were preserved in a dry-ice container and transported to the laboratory for later analyses. We identified twenty algae up to the lowest possible taxonomic level (Littler & Littler, 2000). The species identified were *Amphiroa fragilissima*, *Acanthophora spicifera*, *Bryothamnion triquetrum*, *Caulerpa racemosa*, *Ceramium nitens*, *Chaetomorpha crassa*, *Cladophora vagabunda*, *Dasya antillarum*, *Dictyota caribaea*, *Dictyota menstrualis*, *Gelidiella acerosa*, *Dichotomaria obtusata*, *Galaxaura rugosa*, *Gracilaria spp.*, *Halimeda scabra*, *Hypnea valentiae*, *Jania adhaerens*, *Padina boergesenii*, *Palisada perforata*, and *Tricleocarpa fragilis*.

**Sample analysis:** Carbon and nitrogen were measured from the muscle of the Aristotle’s lantern (echinoids), and from bulk tissues (algae). Once defrosted, calcified algae
samples were washed twice with diluted solutions of HCl at 12 % and 10 % respectively, to remove any excess carbonate (Bunn, Loneragan, & Kempster, 1995). Samples were rinsed with distilled water and dried in an air circulating Shel Lab Oven at 60 °C for 48 hours. After drying, samples were ground to a fine powder using a frequency grinder. Because the acid-wash procedure alters δ¹⁵N results, calcified algae required two samples each: one acid washed for δ¹³C and one without acid-wash for δ¹⁵N. Samples of 0.9-1.3 mg for sea urchin tissue, and 3.9-4.4 mg for algae samples were stored in tin capsules. Measurements of stable isotope ratios were obtained in a continuous flow isotope ratio mass spectrometer at the Laboratory of Stable Isotope Ecology in Tropical Ecosystems, at the University of Miami. Results for δ¹³C and δ¹⁵N were expressed as parts per thousand (%) differences from the standard (δ): Vienna Pee Dee Belemnite (vPDB) for δ¹³C, and atmospheric N² for δ¹⁵N. Carbon and nitrogen samples were analyzed in a dual isotope mode. Ratios were then used to complete equation 1, where \( R_{\text{sample}} \) and \( R_{\text{standard}} \) are the values of \(^{13}\text{C}/^{12}\text{C}\) or \(^{15}\text{N}/^{14}\text{N}\) in the sample and the standard, respectively.

\[
X (\%) = \left( \frac{R_{\text{sample}}}{R_{\text{standard}}} - 1 \right) \times 1000 \quad \text{Eq. 1}
\]

For \textit{E. lucunter} only, we run a Bayesian mixing model and SIBER using SIAR 4.2 (Stable Isotopes Analysis in R) package in the repository Comprehensive R Archive Network (Parnell & Jackson, 2013). This package uses organism isotopic data to fit a model to their dietary habits based upon a Gaussian likelihood and a mixture Dirichlet-distributed prior on the mean. Models were run with 100000 iterations and a burn in of 50000. This routine takes bivariate isotope data in \( x \) and \( y \), and returns the posterior draws for the metric SEA.B (Standard Ellipse Area. Bayesian) for each group. We selected eight algae at every site, except in LQY, where we used seven species, and the percent of contribution of each algae species to the measured isotopic signatures was determined. We run SIBER (Stable Isotope Bayesian Ellipses) to calculate ellipse based metrics of niche width, and determined the existence of any overlap among co-occurring species. SIBER was run twice, once for the hardground biotope (\textit{D. antillarum}, \textit{E. lucunter}, and \textit{E. viridis}), and again for the seagrass biotope (\textit{T. ventricosus}, and \textit{L. variegatus}). We considered an isotopic fractionation across trophic levels of 3.4 ‰ for δ¹⁵N and 1.0 ‰ for δ¹³C (Vander-Zanden & Rasmussen, 2001; Post, 2002; Wing et al., 2008).

Normality and homogeneity of variance were tested by running the Kolmogorov-Smirnov and Cochran tests, respectively (Zar, 2010). Nitrogen data followed the premises of parametric analysis, but the carbon data required a square-root transformation (Box & Cox, 1964). We ran a one-way ANOVA for comparisons of δ¹⁵N and δ¹³C among sea urchins, and a \textit{a posteriori} Tukey-HSD test was used for multiple comparisons of the means. In addition, we ran a One-way ANOVA to analyze spatial differences among isotopic signals of carbon and nitrogen on \textit{E. lucunter} and the seaweed communities. Statistics were performed with \( \alpha < 0.05 \) in R-2.9.13 (R Development Core Team, 2014).

**RESULTS**

The δ¹⁵N signal for the five studied echi- noids of hardground biotopes varied from 5.77 ± 0.25 [mean ± SD] to 8.22 ± 0.36 ‰; whereas the species collected on seagrass biotopes showed values of nitrogen from 6.09 ± 0.79 to 8.02 ± 0.36 ‰ (Fig. 2A). The species \textit{T. ventricosus} and \textit{D. antillarum} showed the highest values of δ¹⁵N. Sea urchins exhibited similar values of δ¹³C varying from -11.6 ± 0.63 to -10.4 ± 0.99 ‰. The echinoid \textit{E. lucunter} displayed the lowest values of carbon, from -15.40 ± 0.76 ‰ (Fig. 2B). Significant differences among species were found for δ¹⁵N and δ¹³C (Table 1). A \textit{a posteriori} analysis displayed differences in δ¹⁵N between the group formed by \textit{E. lucunter}, \textit{E. viridis} and \textit{L. variegatus} and the upper group formed by \textit{D. antillarum} and


T. ventricosus (p< 0.001, Fig 2A). Whereas differences in terms of δ13C were found in E. lucunter with respect to the other four species (p< 0.001, Fig 2B). We found no differences in δ15N signal of E. lucunter among sites, but we did find spatial differences in δ13C (F= 7.410, df= 3, p= 0.001). Those spatial differences were between the sites CGD and TMD (p= 0.001), and between CGD and LQY (p= 0.020).

The overall isotopic signal of δ15N for seaweed communities associated with hardground biotopes varied from 1.7 ± 0.56 (MLN) to 3.3 ± 1.24 ‰ (CGD), whereas the δ13C varied from -15.2 ± 2.51 (LQY) to -13.5 ± 3.58 ‰ (CGD). The lowest nitrogen isotopic signals were obtained in TMD (D. menstrualis: 1.1 ‰) and MLN (G. rugosa: 1.1 ‰); while the highest value was recorded in CGD (B. triquetrum: 4.8 ‰). Both maximum (J. adhaerens: -8.1 ‰) and minimum (D. menstrualis: -8.7 ‰) δ13C values were recorded in CGD (Fig. 3). No differences among sites were detected for overall δ15N (F= 1.300, df= 3, p= 0.301), but we found spatial differences for δ13C (F= 7.410, df= 3, p= 0.001). A posteriori analysis displayed differences between CGD with respect to the other three sites (p< 0.05).

The ellipse-based metrics of niche width analysis -restricted to one locality- found that the hardground biotope species (D. antillarum, E. lucunter, and E. viridis) did not overlap each other (Fig 4A). Similar results were obtained for the co-occurring species of the seagrass biotope; however, the distance between these species was closer than that of the hardground biotope species (Fig 4B). The Bayesian mixing models run for E. lucunter at all four localities found differences in food resources contribution. For instance, the algae D. menstrualis, C. crassa and B. triquetrum dominated in CGD; whereas C. nitens, Gracilaria spp., and D. caribaea represented the main algae species to the diet of E. lucunter at LQY (Table 2). In Culebra Island, no dominance of any particular algae was detected in TMD, where six of the eight species exhibited a similar contribution (Table 2). Although MLN displayed a similar low dominance of species as that of TMD, two algae remained with a mean contribution over 15 %: Dichotomaria obtusata and Dasya antillarum.

**TABLE 1**

Analysis of variance of the isotopic signals of Carbon and Nitrogen among the five sea urchin species

<table>
<thead>
<tr>
<th>Species</th>
<th>df</th>
<th>F</th>
<th>P</th>
<th>df</th>
<th>F</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>δ13C</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Error</td>
<td>20</td>
<td>11.35</td>
<td>0.568</td>
<td>20</td>
<td>9.68</td>
<td>0.48</td>
</tr>
<tr>
<td>Species</td>
<td>4</td>
<td>33.08</td>
<td>&lt; 0.001*</td>
<td>4</td>
<td>15.04</td>
<td>&lt; 0.001*</td>
</tr>
<tr>
<td>SS</td>
<td></td>
<td></td>
<td></td>
<td>SS</td>
<td></td>
<td></td>
</tr>
<tr>
<td>MS</td>
<td></td>
<td></td>
<td></td>
<td>MS</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

* significance.

Fig. 2. δ15N (A) and δ13C (B) values of five echinoids of Puerto Rico (mean ± S.D.).

Fig. 3. Isotopic signal of carbon and nitrogen of the potential food resources of the sea urchin *Echinometra lucunter* (in red) of hardground biotopes at four localities of Puerto Rico Archipelago. Correction factors used for the mixture (echinoid) were 3.4 ‰ for δ^{15}N and 1.0 ‰ for δ^{13}C. The δ^{15}N scale differs between TMD/MLN and CGD/LQY. The sample size was five individuals per species.

**TABLE 2**
Average percentage contribution of seaweeds to the diet of the sea urchin *E. lucunter* at four sites of Puerto Rico Archipelago

<table>
<thead>
<tr>
<th>Seaweed Species</th>
<th>CGD Mean</th>
<th>Credibility Min</th>
<th>Interval Max</th>
<th>LQY Mean</th>
<th>Credibility Min</th>
<th>Interval Max</th>
</tr>
</thead>
<tbody>
<tr>
<td>D. menstrualis</td>
<td>0.24</td>
<td>0.08</td>
<td>0.41</td>
<td>C. nitens</td>
<td>0.22</td>
<td>0.03</td>
</tr>
<tr>
<td>C. crassa</td>
<td>0.21</td>
<td>0.06</td>
<td>0.40</td>
<td>Gracilaria spp.</td>
<td>0.20</td>
<td>0.01</td>
</tr>
<tr>
<td>B. triquetrimum</td>
<td>0.20</td>
<td>0.01</td>
<td>0.38</td>
<td>D. caribaea</td>
<td>0.17</td>
<td>0</td>
</tr>
<tr>
<td>P. boegesennii</td>
<td>0.11</td>
<td>0</td>
<td>0.24</td>
<td>A. spicifera</td>
<td>0.12</td>
<td>0</td>
</tr>
<tr>
<td>A. fragilissima</td>
<td>0.07</td>
<td>0</td>
<td>0.16</td>
<td>P. perforata</td>
<td>0.11</td>
<td>0</td>
</tr>
<tr>
<td>G. acerosa</td>
<td>0.07</td>
<td>0</td>
<td>0.19</td>
<td>C. vagabunda</td>
<td>0.11</td>
<td>0</td>
</tr>
<tr>
<td>G. rugosa</td>
<td>0.07</td>
<td>0</td>
<td>0.18</td>
<td>G. rugosa</td>
<td>0.07</td>
<td>0</td>
</tr>
<tr>
<td>J. adhaerens</td>
<td>0.04</td>
<td>0</td>
<td>0.10</td>
<td></td>
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<td></td>
</tr>
</tbody>
</table>

**TMD**

<table>
<thead>
<tr>
<th>Seaweed Species</th>
<th>Mean</th>
<th>Credibility Min</th>
<th>Interval Max</th>
<th>LQY Mean</th>
<th>Credibility Min</th>
<th>Interval Max</th>
</tr>
</thead>
<tbody>
<tr>
<td>H. scabra</td>
<td>0.16</td>
<td>0.01</td>
<td>0.28</td>
<td>D. obtusata</td>
<td>0.18</td>
<td>0.05</td>
</tr>
<tr>
<td>G. rugosa</td>
<td>0.14</td>
<td>0.01</td>
<td>0.26</td>
<td>D. antillarum</td>
<td>0.15</td>
<td>0.01</td>
</tr>
<tr>
<td>D. mensuralis</td>
<td>0.14</td>
<td>0.001</td>
<td>0.26</td>
<td>G. rugosa</td>
<td>0.14</td>
<td>0.01</td>
</tr>
<tr>
<td>H. valentiae</td>
<td>0.14</td>
<td>0</td>
<td>0.27</td>
<td>D. caribaea</td>
<td>0.12</td>
<td>0</td>
</tr>
<tr>
<td>T. fibrillos</td>
<td>0.14</td>
<td>0.001</td>
<td>0.26</td>
<td>C. nitens</td>
<td>0.12</td>
<td>0.004</td>
</tr>
<tr>
<td>G. acerosa</td>
<td>0.13</td>
<td>0.002</td>
<td>0.25</td>
<td>D. mensuralis</td>
<td>0.11</td>
<td>0</td>
</tr>
<tr>
<td>A. spicifera</td>
<td>0.09</td>
<td>0</td>
<td>0.20</td>
<td>H. valentiae</td>
<td>0.10</td>
<td>0</td>
</tr>
<tr>
<td>C. racemosa</td>
<td>0.06</td>
<td>0</td>
<td>0.17</td>
<td>G. acerosa</td>
<td>0.09</td>
<td>0</td>
</tr>
</tbody>
</table>

Sites are Cerro Gordo (CGD), Luquillo (LQY), Tamarindo (TMD), and Melones (MLN).
DISCUSSION

There is often a high degree of spatial variability in shallow-water ecosystems. Both nitrogen and carbon elemental concentrations may vary due to abiotic and biotic factors, including atmospheric deposition, volatilization, microbial processing, terrestrial runoff, etc. (Van-Dover et al., 1992; Nixon et al., 1996). Anthropogenic enrichment may alter marine benthic food webs, increasing the ratios of heavy isotopes in primary producers (McGlathery, 2001; Tewfik, Rasmussen, & McCann, 2005). In fact, the presence of elevated values of $\delta^{15}N$ has been shown to be related with local NO$_3$ enrichment (Clark & Wilcock, 2000; Cloern, 2001). In general, our seaweed communities exhibited no spatial differences in nitrogen, which indicates a relative homogeneity in $\delta^{15}N$ among sites. This is probably due to their geographical proximity. In contrast, spatial differences among seaweed communities in terms of $\delta^{13}C$ might be related to community composition, metabolic species-specific characteristics, or to local geomorphologic features. The CGD site exhibited differences with respect to the other three sites. This difference may be linked to the existence of a mangrove lagoon canal located approximately 200 m away. This is important because mangrove systems provide nutrient inputs, mainly carbon, to nearshore waters (Zieman, Macko, & Mills, 1984; Rezende, Lacerda, Ovall, Silva & Martinelli, 1990).

Food resources leave tracks within consumers throughout food web (Phillips, 2012), and usually one or two sources may contribute 50% or more of the diet of a given consumer (Rodriguez, 2003; Tomas et al., 2006). Mixing models indicated spatial differences among sites in the composition of algal food assimilated by the sea urchin *E. lucunter*. No single algal species exhibited high dominance in the diet of this sea urchin in TMD and MLN.

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**Fig. 4.** Results of the ellipses based metrics of niche width for two echinoids groups based on habitat preferences. **A:** represents the three species inhabit in hardgrounds whereas. **B:** represents the species of seagrass meadows. Circles represent the potential niche of each species.
However, some algae reported mean dietary contribution greater than 20% in CGD and LQY, but never as high as 30 or 40% for a single species. These results contrast with other studies where some algae species may compromise up to 60% of urchin diets (Rodríguez, 2003; Prado et al., 2010; Cabanillas-Terán et al., 2016). Although contrasting, trophic plasticity is widespread in nature, and may explain the apparent difference in urchin diets (Pinnegar & Polunin, 2000; Wing et al., 2008).

The existence of spatial differences reflects the feeding plasticity of *E. lucunter* and its ability to use a wide range of food resources. The fact that the Culebra sites showed a lower algal contribution in sea urchin diets might be related to local seaweed assemblages than to feeding preferences. A generalist feeding behavior is very common in sea urchins, and *E. lucunter* is not an exception (Hendler et al., 1995; Lawrence et al., 2013). The lack of data on seaweed communities did not allow us to quantify algal abundances or establish a relationship between seaweed abundance and its magnitude of contribution to the diet of *E. lucunter*. We do not suspect that the reason why the sea urchin diet is distributed among so many algae types is due simply to the low species number (8) included in this two-isotope mixing model. Previous studies have stated that those resources, whose isotopic signals most resemble those of the sea urchin, are in general the most important contributors to the diet of a given species (Szeplanski, Ben-David, & Van Ballenberghe, 1999; Ben-David & Shell, 2001). There is a possibility that *E. lucunter* may be selectively feeding on a few algal species, and different combinations could produce the same isotopic signal; nevertheless, further studies will need to focus on isotopic markers to prove this diversity in the diet of *E. lucunter*.

Co-occurring species coexist due to differences in feeding behavior and internal assimilation mechanisms (Vanderklift et al., 2006; Wolf et al., 2009). Differences in δ¹³C detected among the five echinoids of two different biotopes may suggest that *E. lucunter* uses different sources of carbon with respect to the other two species of the hardground biotope (Fig. 4A), whereas *D. antillarum* and *E. viridis* coincided in terms of δ¹³C. However, the lack of overlap between both species suggests the use of different sources of nitrogen and a higher trophic position for *D. antillarum*, which may be more related to an omnivorous strategy. A similar case of niche partitioning was discussed for three sympatric sea urchins in Australia (Vanderklift et al., 2006).

The other two co-occurring species within seagrass bed habitats also exhibited no niche overlap (Fig 4B). However, both echinoids coincided in terms of δ¹³C, which may indicate the use of similar food resources. A higher trophic position of *T. ventricosus*, however, could indicate the use of other food items, δ¹⁵N-enriched. These two species are common dwellers of seagrass meadows. There, *T. ventricosus* usually grazes on *Thalassia testudinum* leaves, but also ingests small invertebrates; whereas *L. variegatus* feeds on *T. testudinum*, detritus, and some epiphytes (Keller, 1983; Barrios & Reyes, 2009). The isotopic signal of δ¹⁵N for *T. ventricosus* was also in consonance with its omnivorous behavior, and relatively far from the isotopic signature of *T. testudinum* leaves (unpublished data). A stomach content analysis on *T. ventricosus* found bryozoans, ascidians, and hydrozoans (Barrios & Reyes, 2009). Thus, δ¹⁵N in *T. ventricosus* was similar with the isotopic signal of the omnivorous sea urchins *Centrostephanus tenuispinus*, *Phyllophorton irregularis*, *Strongylocentrotus pallidus* and *Arbacia lixula* (Vanderklift et al., 2006; Wangensteen et al., 2011). Nevertheless, the Mediterranean echinoid *Paracentrotus lividus*, considered an important grazer of the vascular plant, *Posidonia oceanica*, reported values of nitrogen between 9-10%. This particularly high value of δ¹⁵N, which could be considered too high for a grazer, was due to the existence of an epiphyte community growing on *Posidonia* leaves that covers part of the carbon and nitrogen requirements in the diet of the echinoid *P. lividus* (Tomas et al., 2006). Therefore, the ingestion of small invertebrates by *T. ventricosus* may be not an accidental
intake, but an important source of nitrogen that gives this sea urchin a higher trophic position in seagrass biotopes.

The high δ^{15}N values found in *D. antillarum* contrasts with its feeding preferences (Rodriguez-Barreras et al., 2015). The sea urchin *D. antillarum* has considered primarily a generalist herbivore that feeds principally upon algae (Hendler et al., 1995). However, this sea urchin also ingests invertebrates (Serafy, 1979). Stomach content analyses have revealed the presence of sponges, hydroids, bryozoans, nematodes, rotifers, gastropods, bivalves, and copepods. These invertebrates may represent up to 32 % of the gut species-richness index (Herrera-López et al, 2004; Hernández et al., 2006). Indeed, some authors proposed *Diadema* as an omnivore (Rotjand & Lewis, 2008), but the idea has met resistance. The presence of invertebrates within the *Diadema* diet has considered a non-selective behavior, due only to accidental scraping off, but never an important component of the sea urchin diet. The high value of δ^{15}N displayed for *D. antillarum*, similar to *T. ventricosus*, suggests that both sea urchins occupy similar trophic levels in their respective habitats, and support an omnivorous feeding behavior for *D. antillarum*. However, this tentative statement requires further studies to determine the main food resources, including invertebrates.

To conclude, a Bayesian mixing model indicates feeding plasticity of *E. lucunter*, using multiple food algal resources depending on local availability. Similarities in δ^{15}N between *D. antillarum* and *T. ventricosus* may hint at a similar trophic level for both species (omnivorous) which is widely accepted for *T. ventricosus*, but not for *D. antillarum*, often considered a generalist herbivore species. Thus, the lack of overlap among species in the two biotopes seems to indicate a resource partitioning strategy to avoid niche competition among co-occurring species.

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**RESUMEN**

Relaciones tróficas entre los erizos de mar Caribe. Las especies *Echinometra lucunter, Echinometra viridis, Lytechinus variegatus, Tripneustes ventricosus*, and *Diadema antillarum* son los erizos de mar más comunes en los hábitat litorales del Caribe. Los erizos de mar *T. ventricosus* y *L. variegatus* habitan generalmente los pastos marinos mientras que las otras tres especies se encuentran asociadas a sustratos rocosos. Los hábitos alimentarios de estas especies han sido bien documentados y son reconocidos como herbívoros – omnívoros; sin embargo, pocas de estas especies han sido caracterizadas isotópicamente. Utilizamos los isótopos estables para caracterizar estas cinco especies de erizos y establecer las posiciones tróficas para las especies que cohabitan los mismos ecosistemas. También cuantificamos la contribución de los recursos alimentarios para *E. lucunter*. Los erizos *T. ventricosus* y *D. antillarum* mostraron los mayores valores de δ^{15}N y valores similares de δ^{13}C que variaron desde -11.6 ± 0.63 a -10.4 ± 0.99 %; donde el erizo *E. lucunter* mostró los valores más negativos con -15.40 ± 0.76 %. Las comunidades de algas no mostraron diferencias en valores promedio de δ^{15}N (F= 1.300, df= 3, p= 0.301), pero sí mostraron variaciones en los valores de δ^{13}C (F= 7.410, df= 3, p= 0.001). Los estudios de amplitud de elipses de nicho determinaron que las especies de los biotopos rocosos (*D. antillarum, E. lucunter* y *E. viridis*) no mostraron solapamiento de nicho. Similar resultado también se encontró en las especies de erizos que habitan en los pastos marinos. Sin embargo, la distancia entre estas dos especies fue menor respecto a la distancia entre las especies de erizos que habitan en los sustratos rocosos. Nuestros resultados muestran que las especies que habitan en los pastos marinos mostraron valores más elevados de δ^{13}C en comparación con las especies de los sustratos rocosos. No se encontraron diferencias espaciales para *E. lucunter* en δ^{15}N, pero sí en δ^{13}C. Los modelos de mezcla bayesianos demuestran la plasticidad alimentaria de *E. lucunter*, especie capaz de utilizar múltiples recursos algaes dependiendo de la disponibilidad por
sitio. Semejanzas en δ¹⁵N entre D. antillarum y T. ventricosus parecen indicar similitudes tróficas entre ambas especies. Si bien T. ventricosus es reconocido como omnívoro, D. antillarum siempre ha sido considerado un herbívoro generalista. Finalmente, la falta de solapamiento entre las especies en los dos biotopos parece indicar una estrategia de partición de recursos para evitar la competencia de nicho entre especies concurrentes.

**Palabras clave:** isótopos estables, ecología trófica, modelos de mezcla, erizos de mar, Puerto Rico.

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